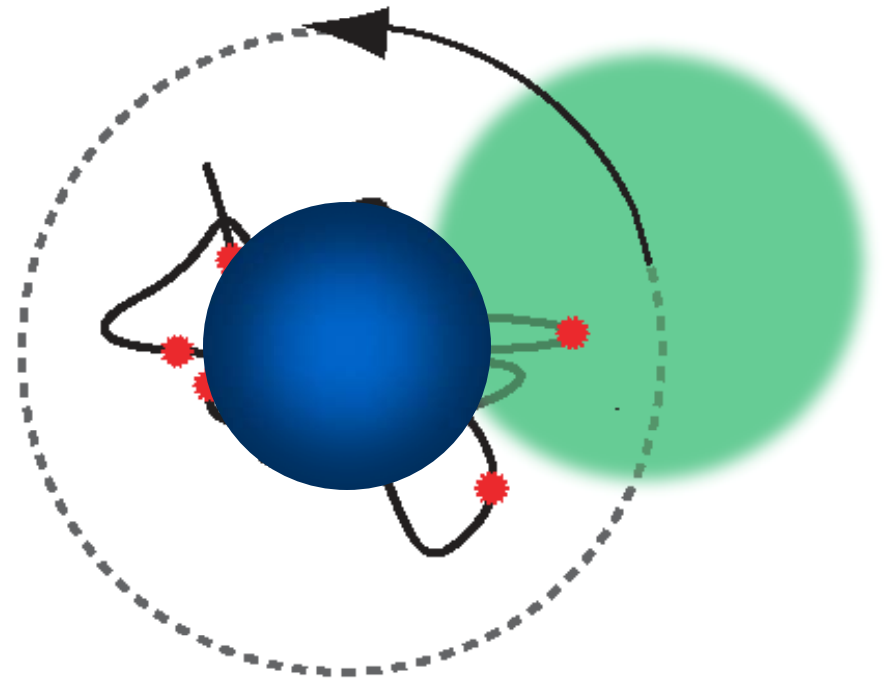
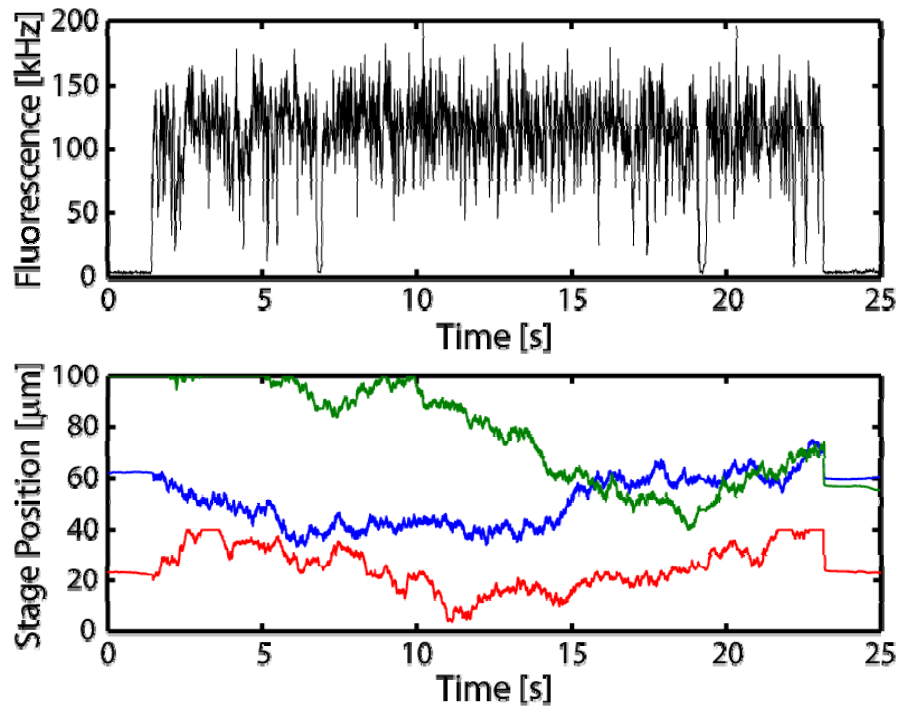


Tracking Fluorescence Correlation Spectroscopy of individual biomolecules

Kevin McHale¹, Andrew Berglund², Mike Zhang, Charles Limouse, Chandra Raman³, Hideo Mabuchi
Edward L. Ginzton Laboratory, Stanford University

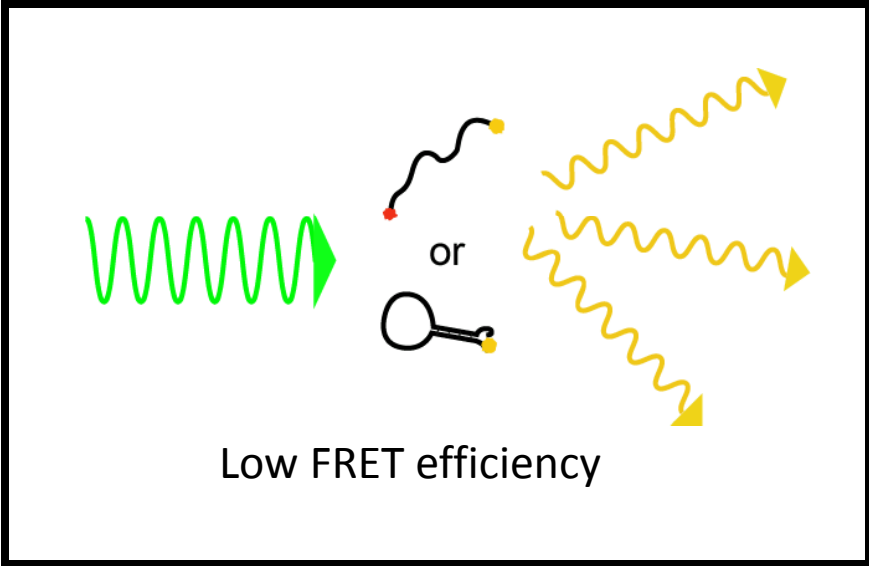
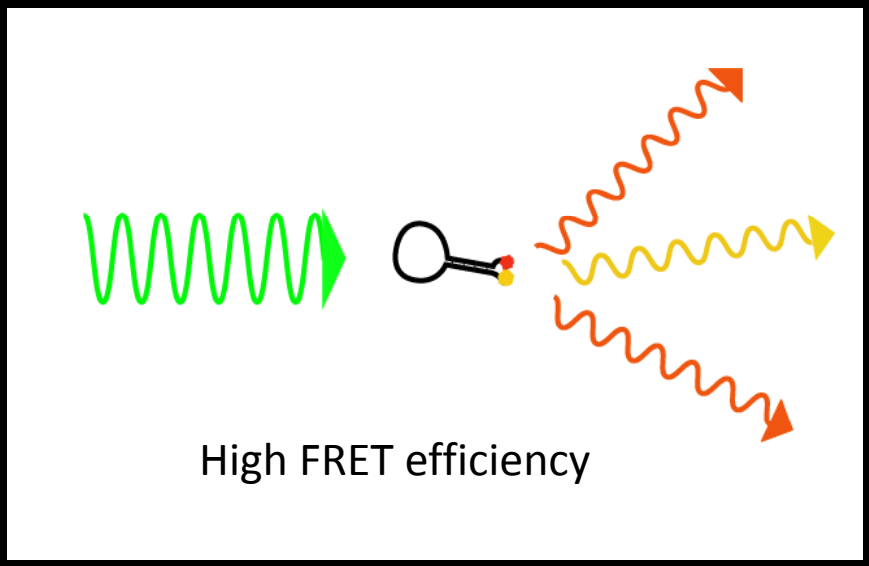
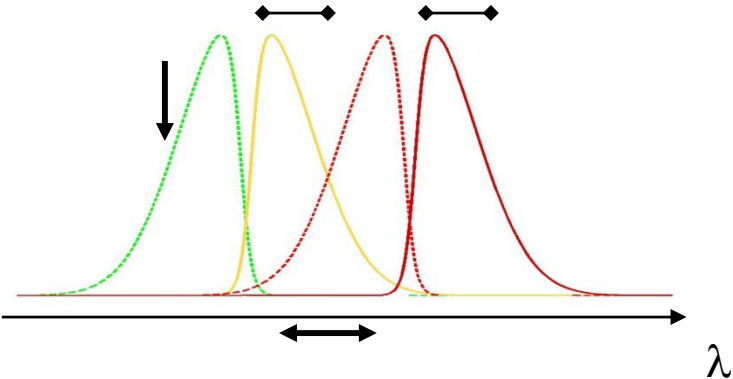
¹Chemical Physics Laboratory, NIDDK, NIH ²CNST Nanofabrication lab, NIST ³Physics, Georgia Institute of Technology



ARO, NDSEG, NSF, NIGMS

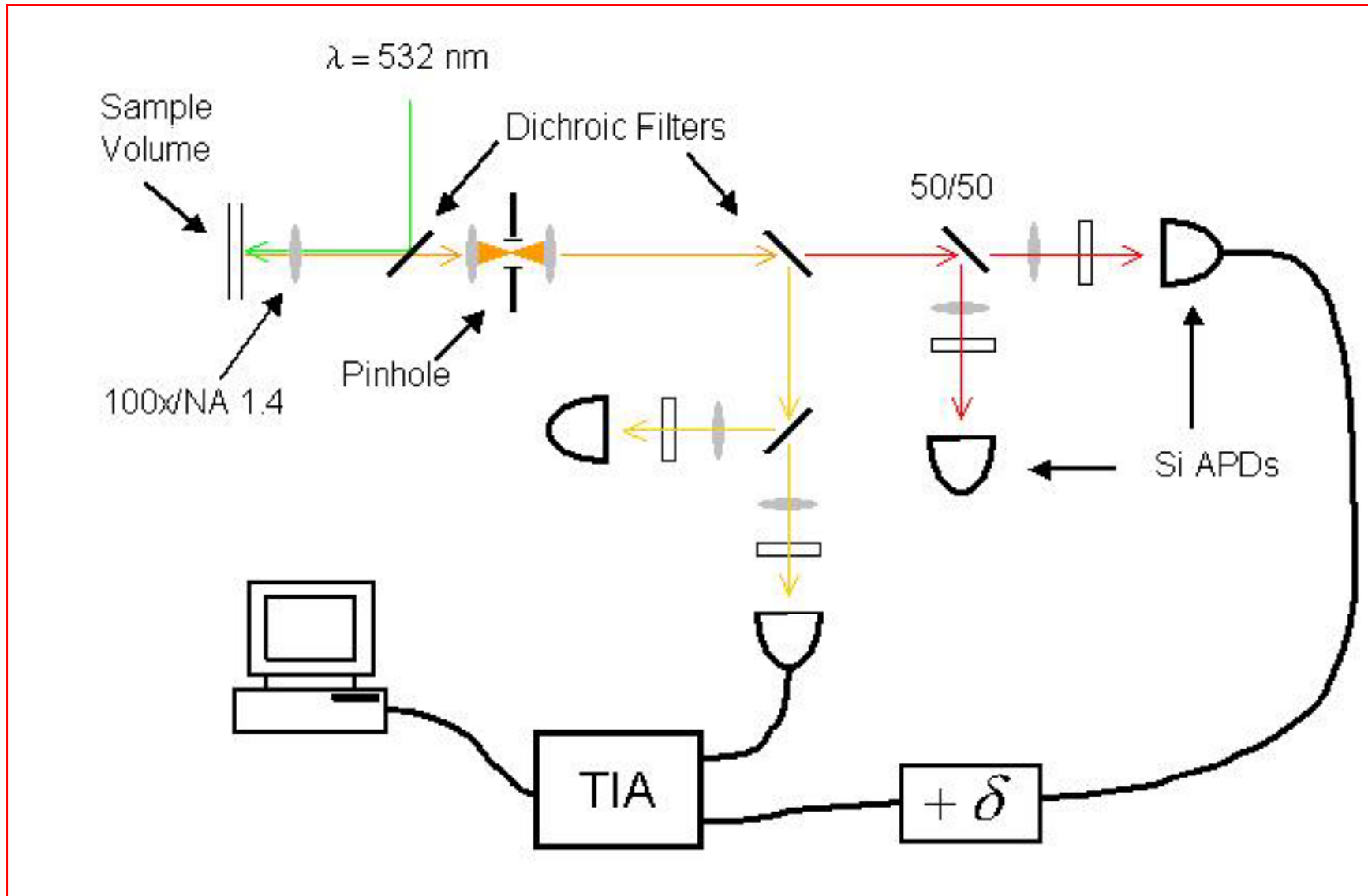
Fluorescence resonance energy transfer (FRET)

- single DNA strands labeled with fluorescent dye molecules
- Position, orientation and spectral overlap determine FRET efficiency

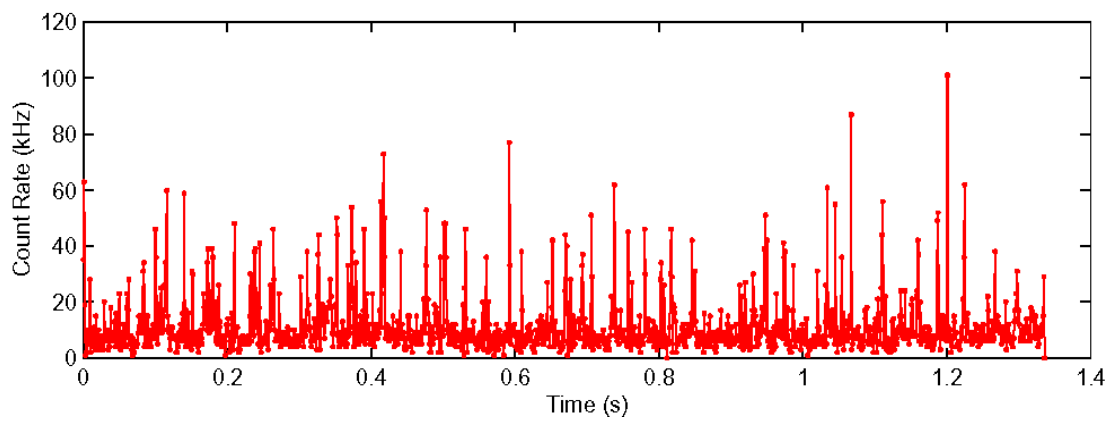
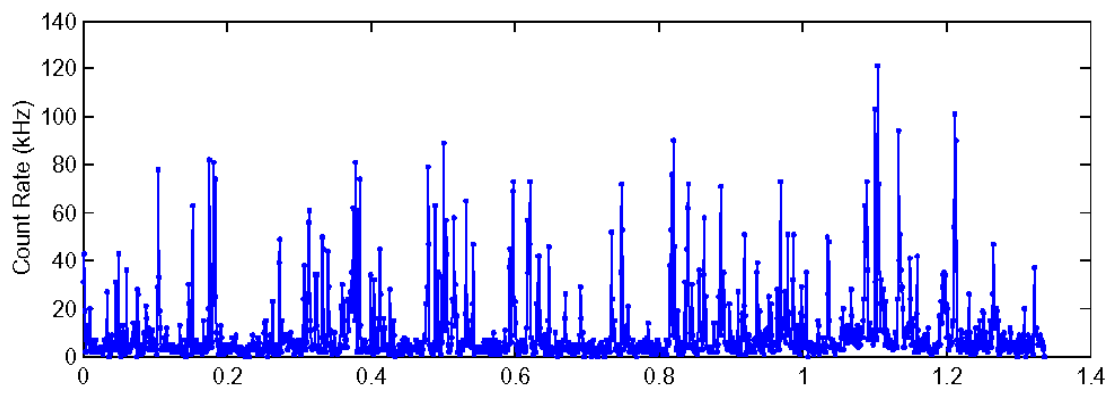
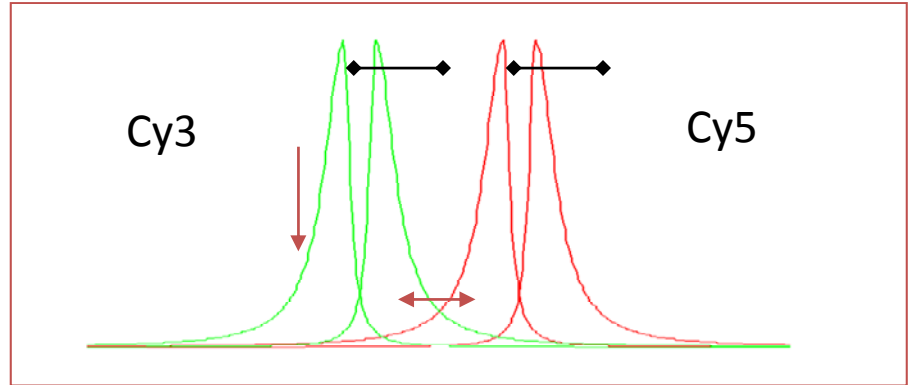
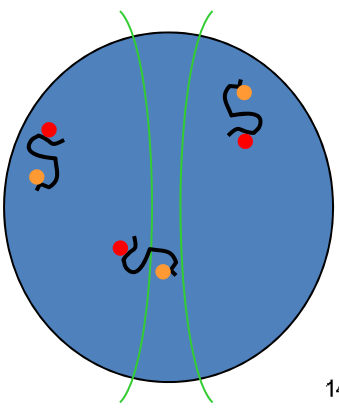


Single-pair FRET measurements on dye-labeled DNA

S. Chu et al, S. Weiss et al



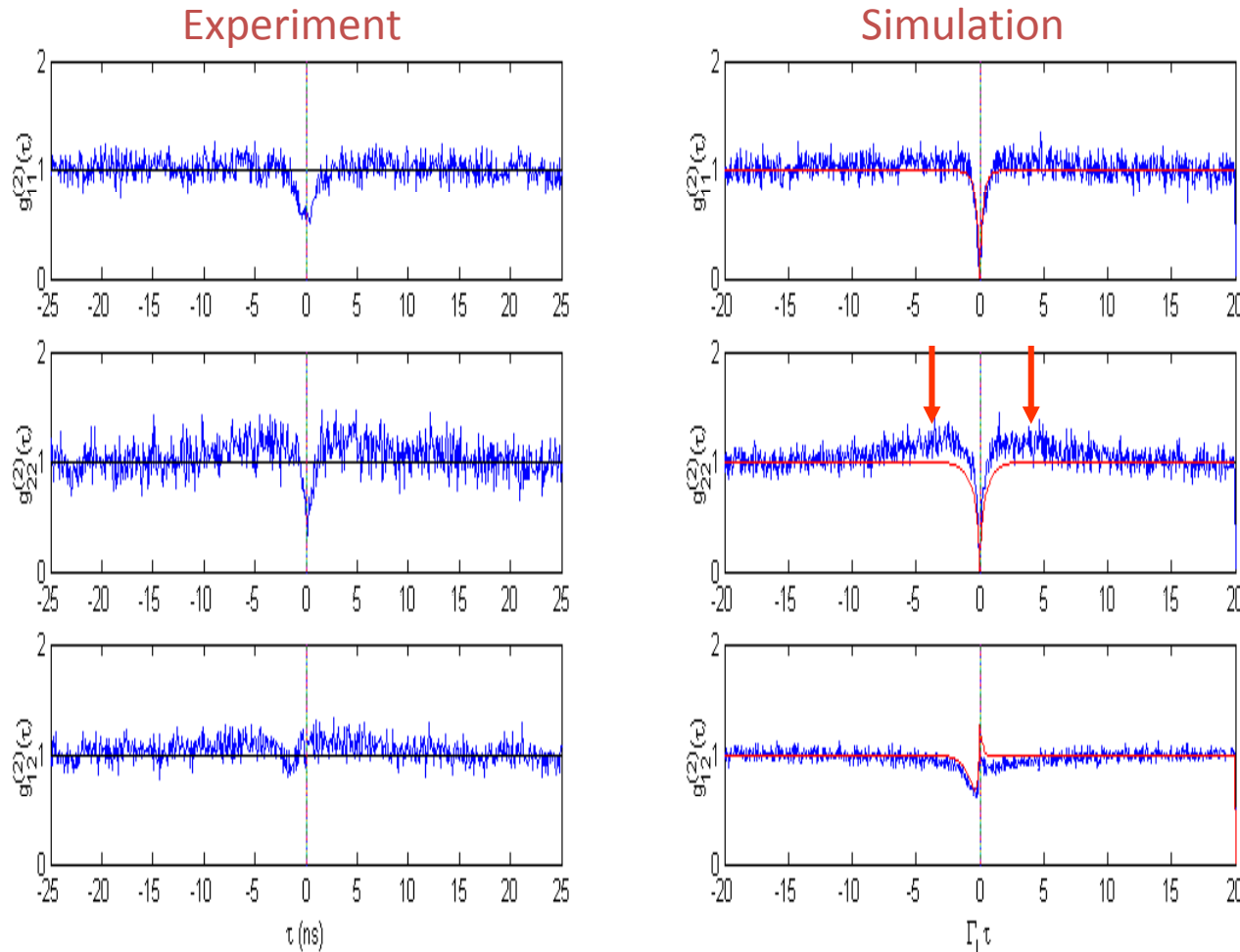
Single-pair FRET measurements on dye-labeled DNA



fluor. ratio
↓
FRET efficiency
↓
relative position, orientation of dyes

Photon statistics and dynamics of FRET

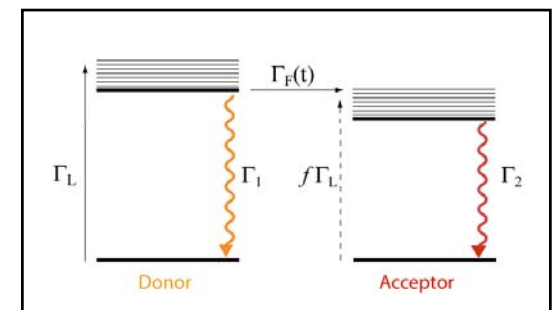
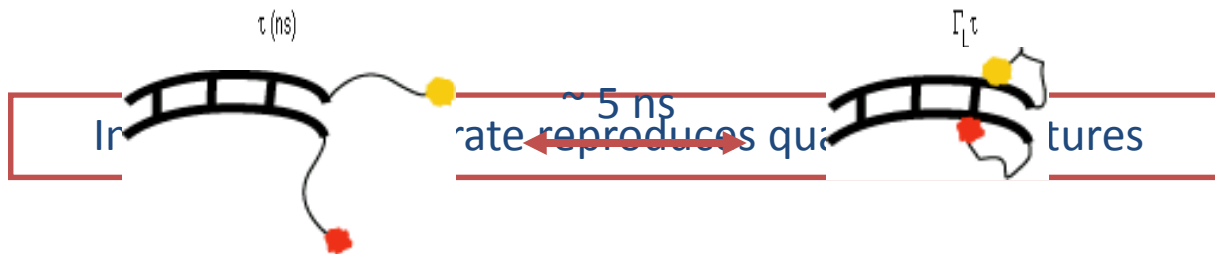
A. J. Berglund, A. C. Doherty and HM, Phys. Rev. Lett. **89**, 068101 (2002)



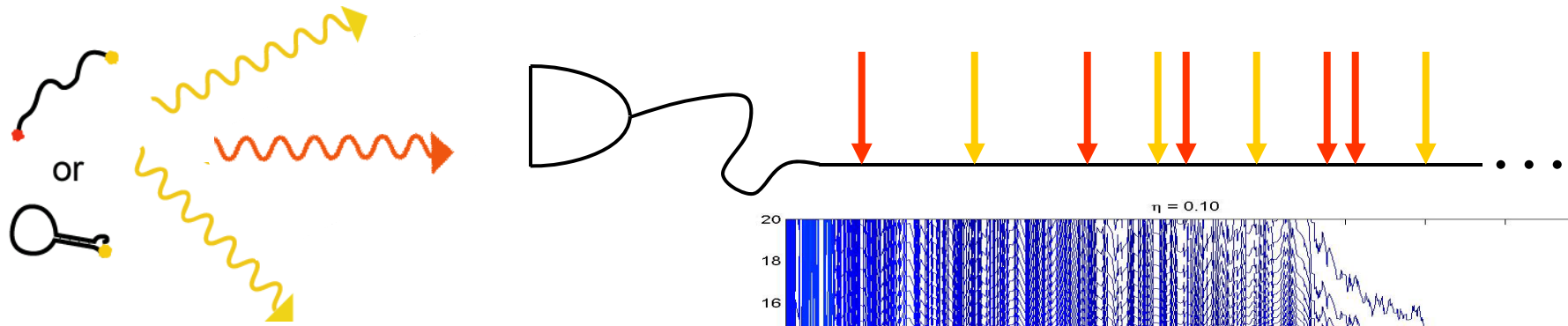
Donor autocorrelation

Acceptor autocorrelation

Cross-correlation

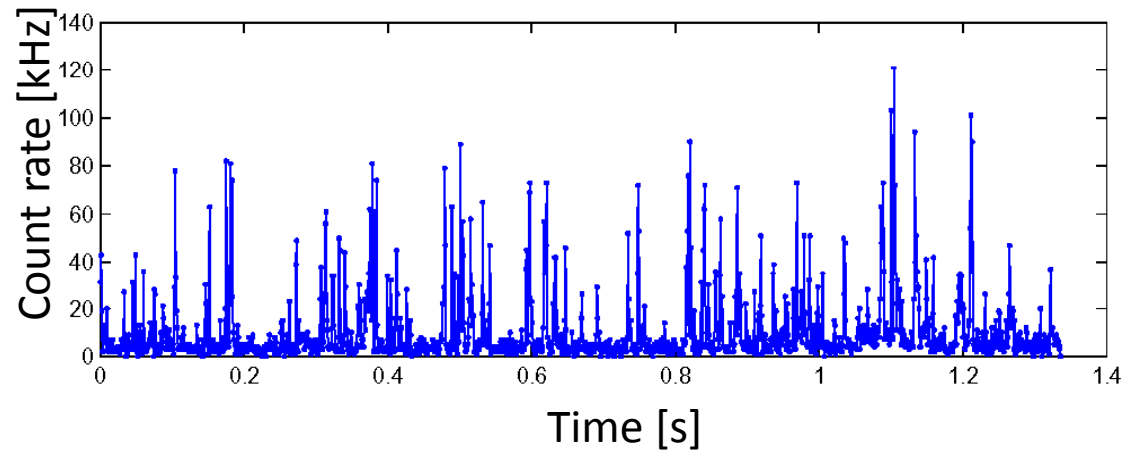
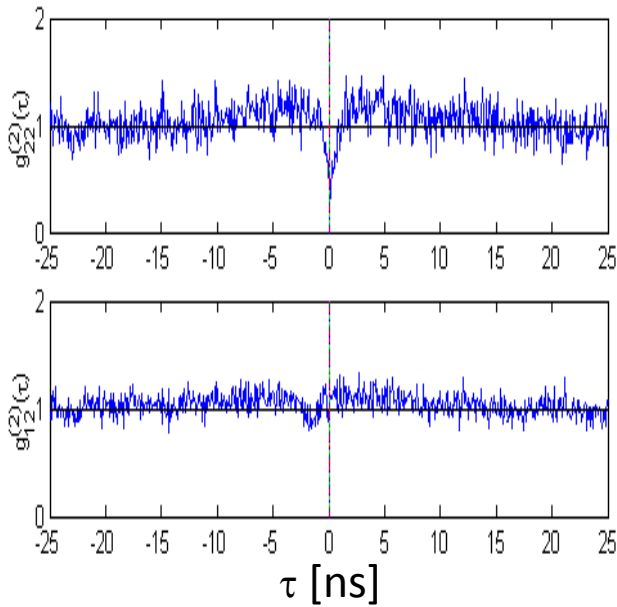
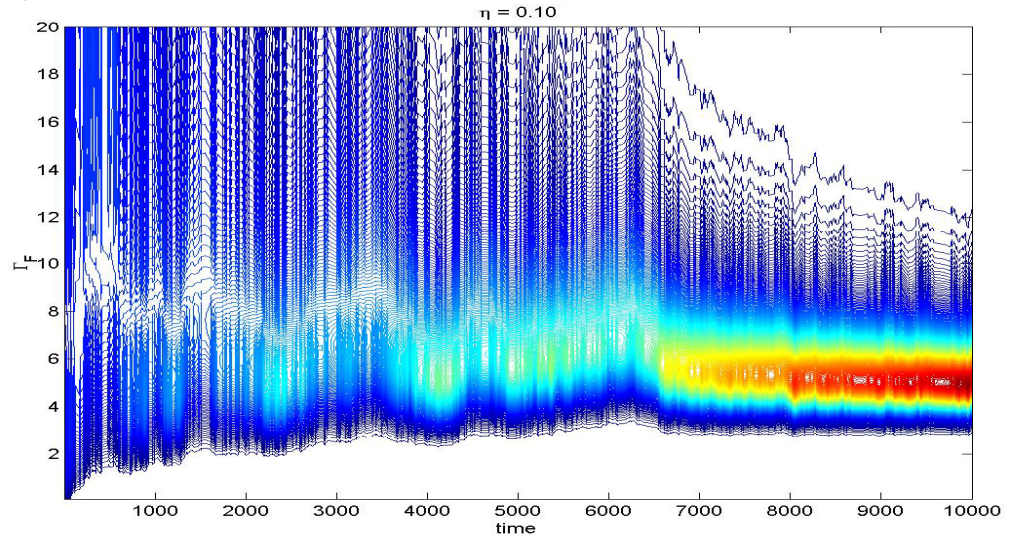


Photon-stream filtering: dynamics vs. shot-noise



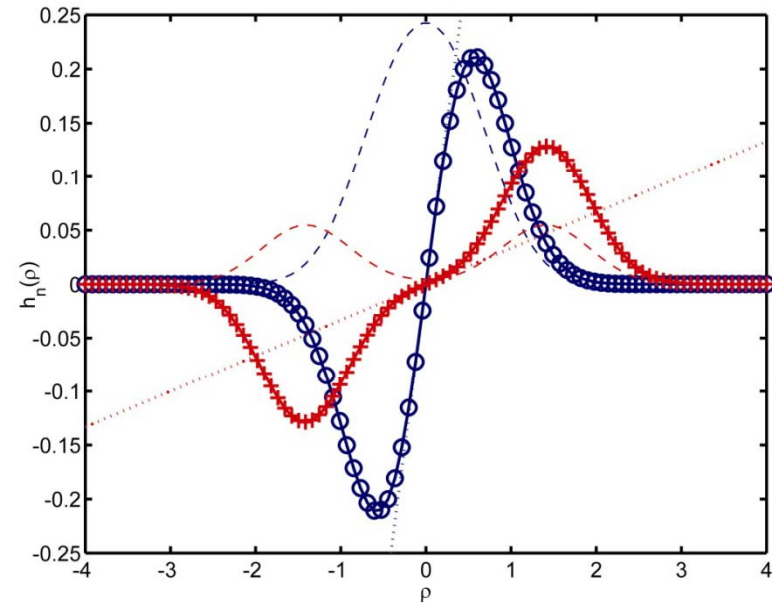
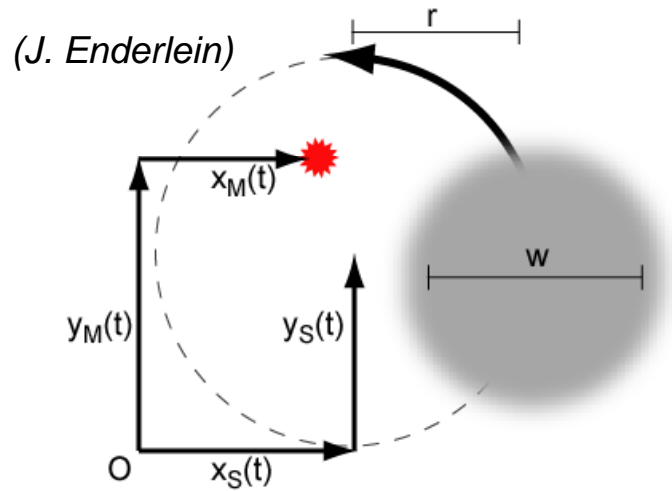
$$p(\vec{\theta} | \Xi[0, t]) = q(\Xi(t) | \vec{\theta}) p(\vec{\theta} | \Xi[0, t])$$

$$d\vec{\theta}_t = \mathcal{A}_t \vec{\theta}_t dt + \mathcal{K}_t [d\Xi_t - C\vec{\theta}_t dt]$$

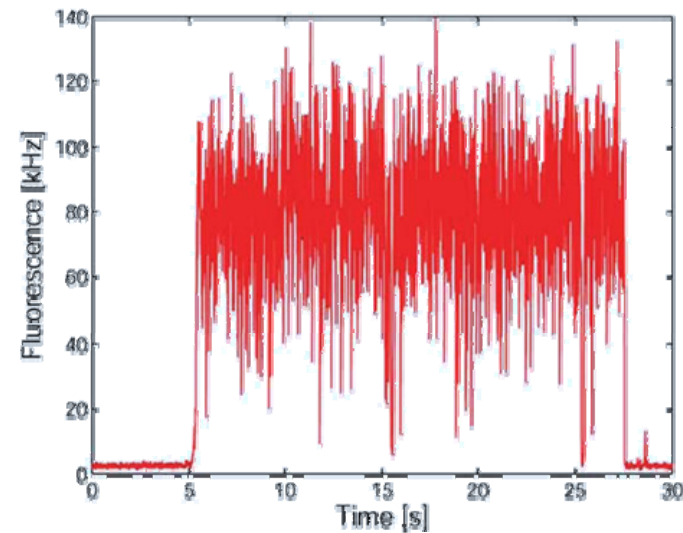
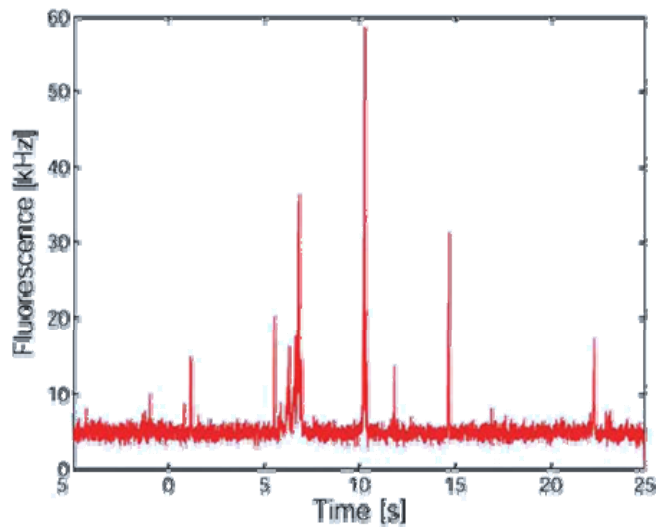


Fluorescence-modulation particle tracking (2D)

A. J. Berglund and HM, Appl. Phys. B **78**, 653 (2004)

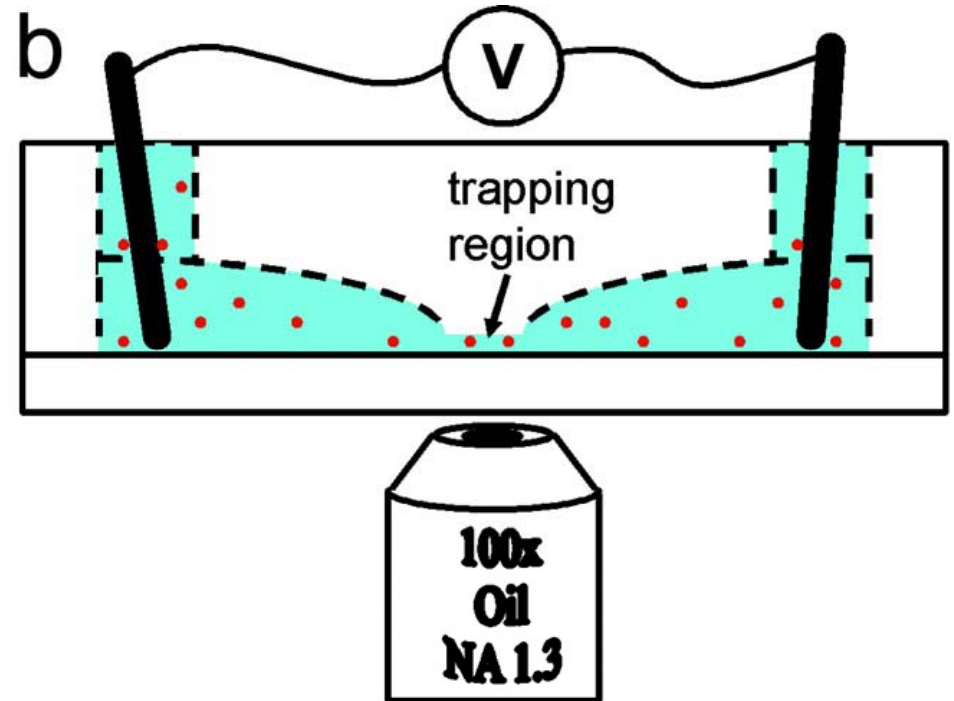
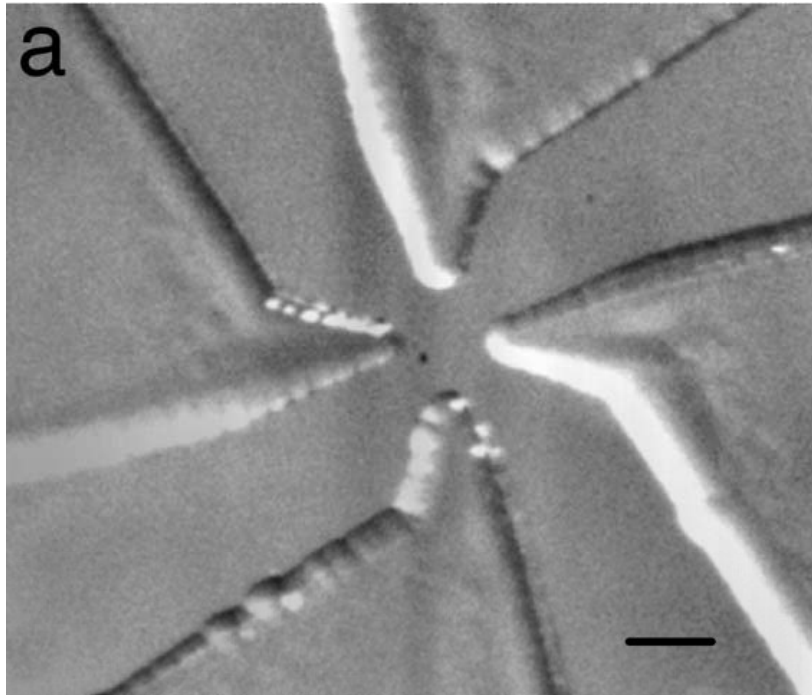


Near axis of rotation the demodulated fluorescence provides a linear error signal

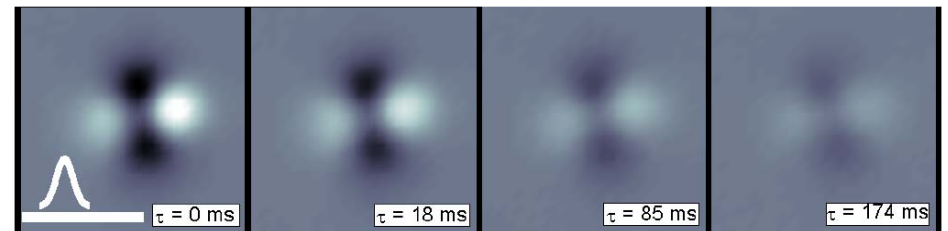


Cohen & Moerner: anti-Brownian electrokinetic trap

A. E. Cohen and W.E. Moerner, Proc. Natl. Acad. Sci. U.S.A. **103**, 4362 (2006)

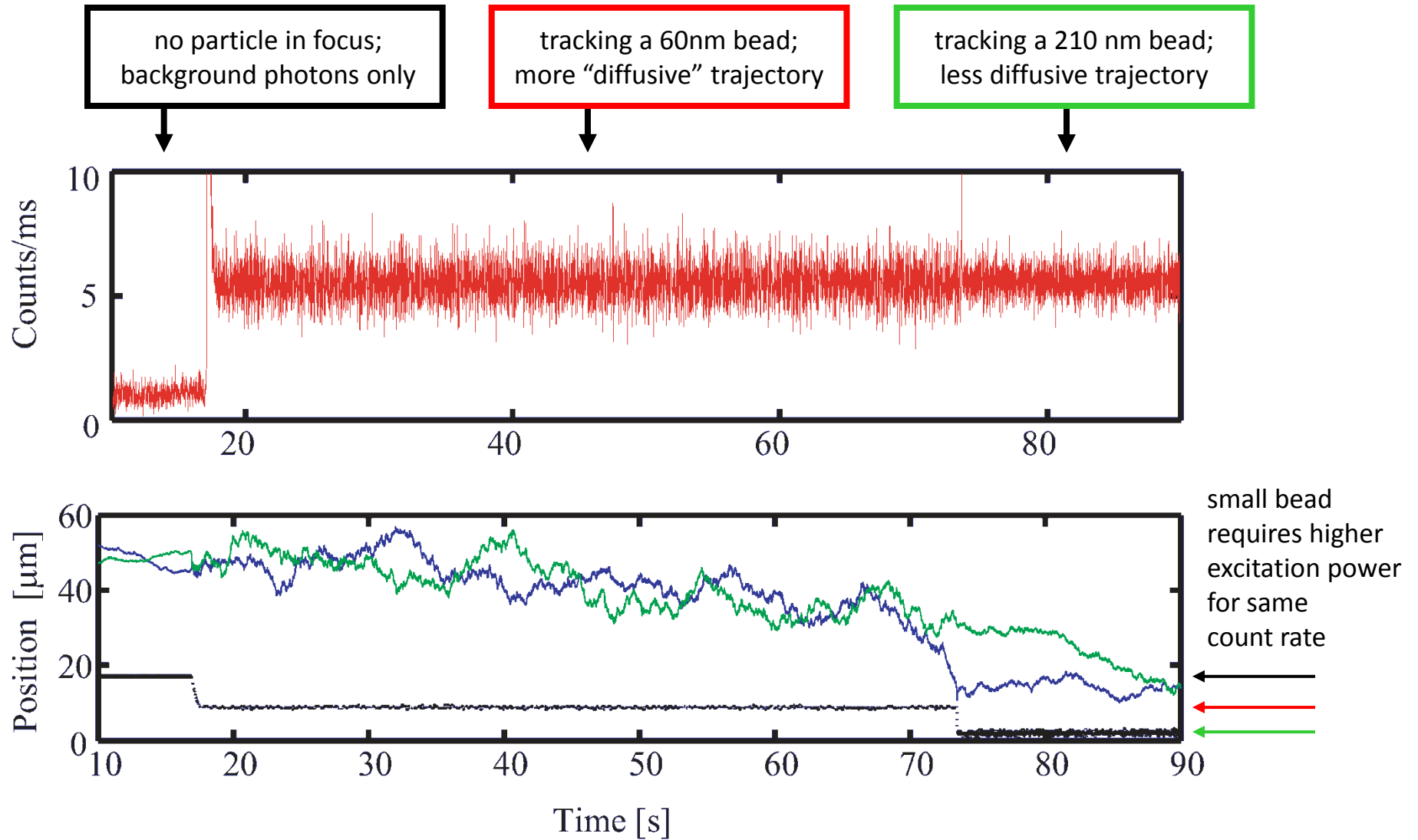


A. E. Cohen and W. E. Moerner, Phys. Rev. Lett. **98**, 116001 (2007)



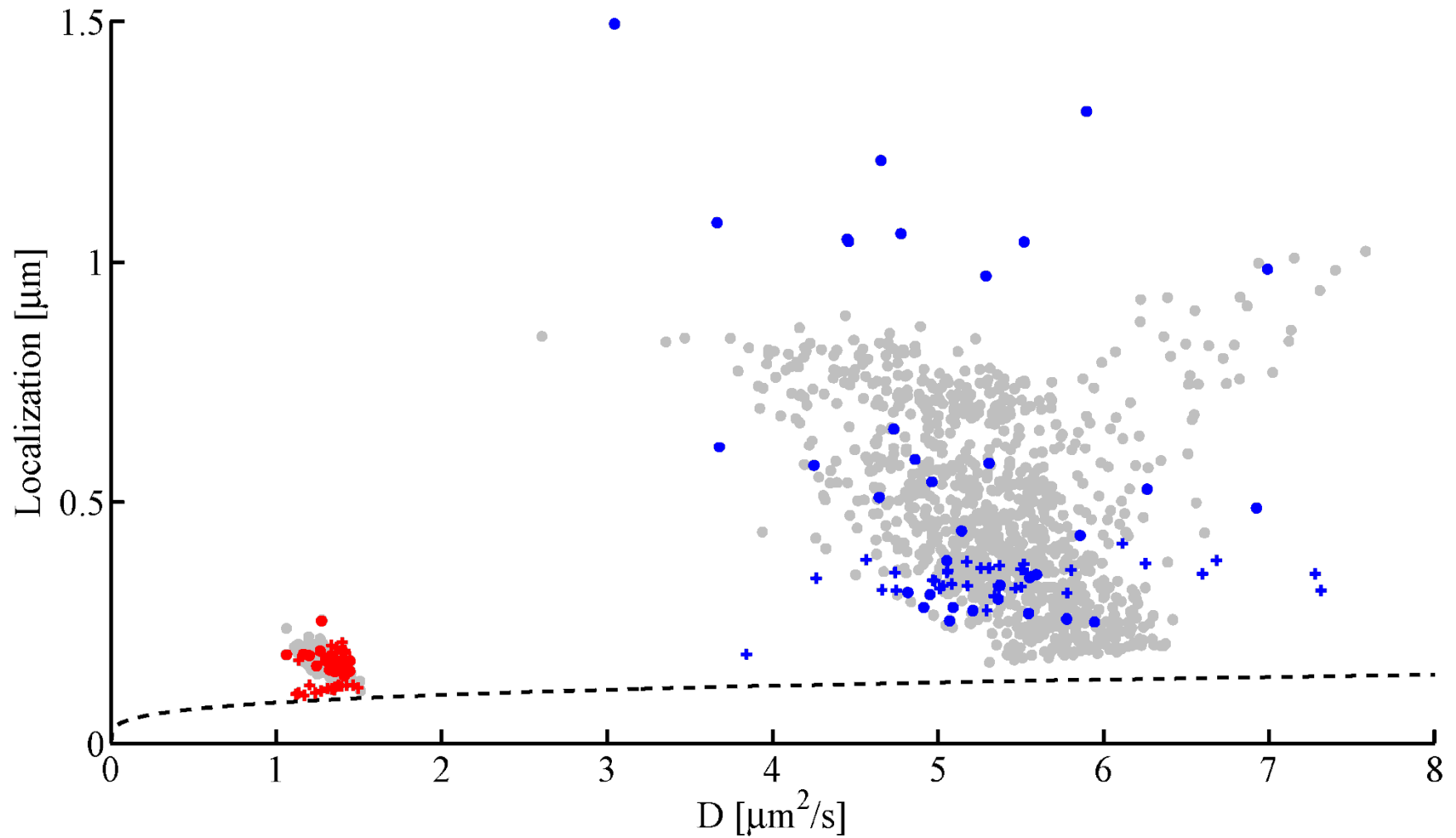
Tracking data channel(s)

A. J. Berglund, K. McHale and HM, Opt. Lett. **32**, 145 (2007)



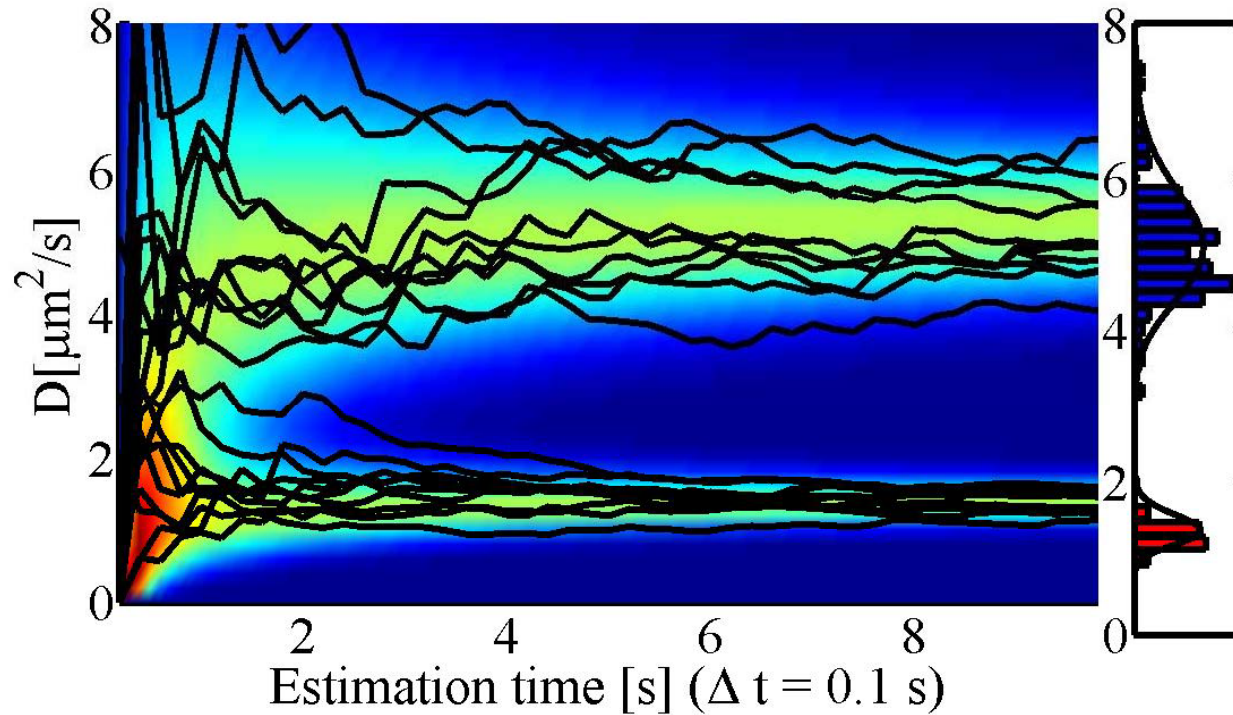
Tracking performance near the shotnoise limit (2D)

A. J. Berglund, K. McHale and HM, Opt. Lett. **32**, 145 (2007)

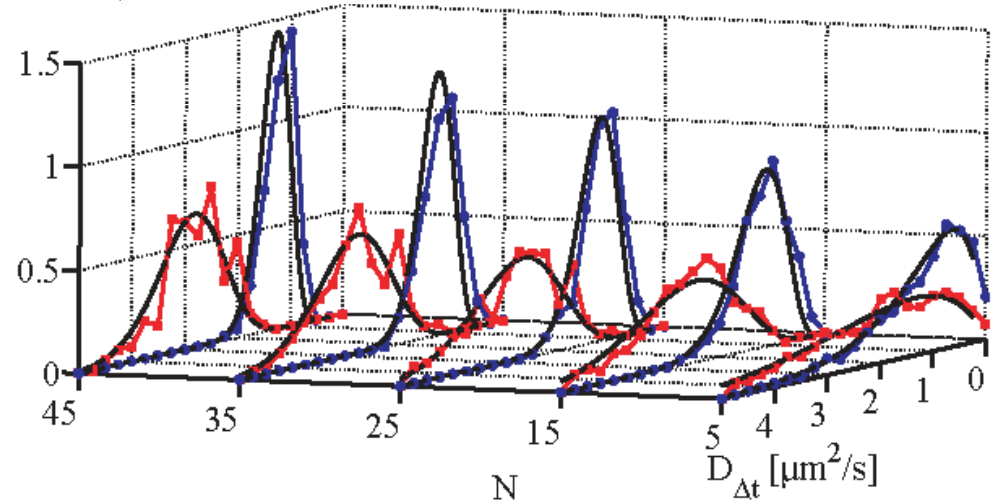
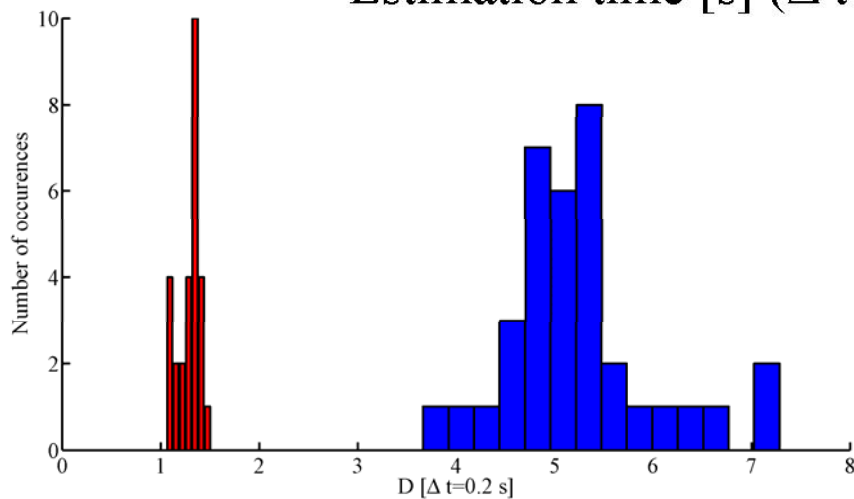


Fast classification of individual nano-particles (beads)

A. J. Berglund, K. McHale and HM, Opt. Express **15**, 7752 (2007)



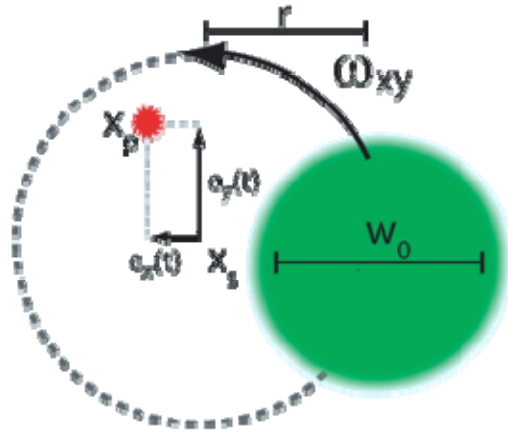
90% confidence
with only ~600
photons!



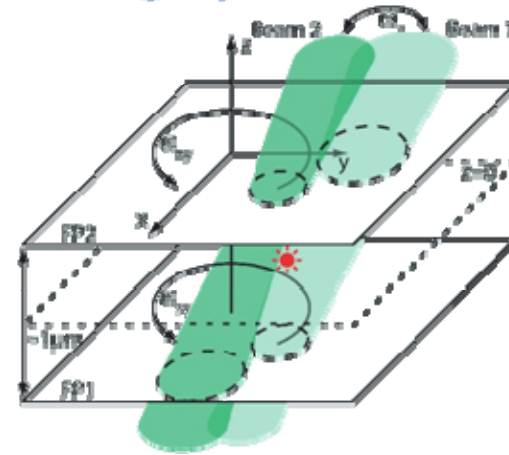
3D Feedback tracking microscopy

Kevin McHale, A. J. Berglund and HM, Nano Lett. **7**, 3535 (2007)

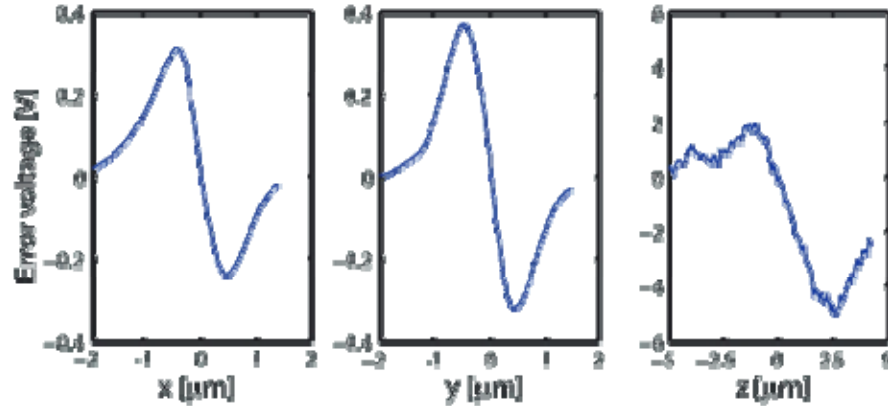
2-D sensing by beam rotation



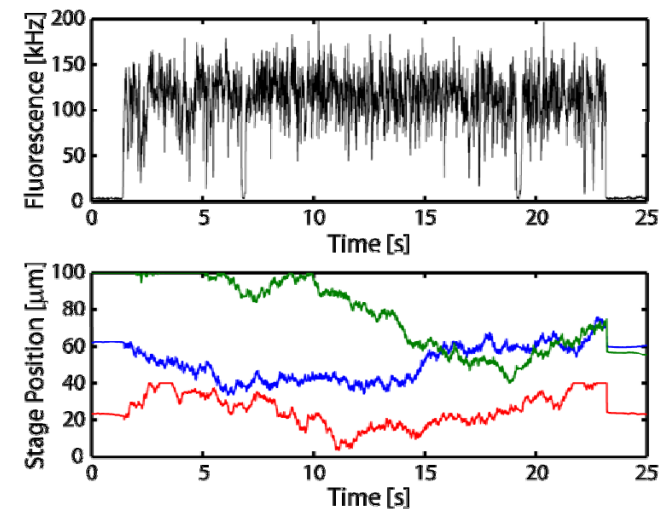
3-D sensing by beam switching



Position error estimates

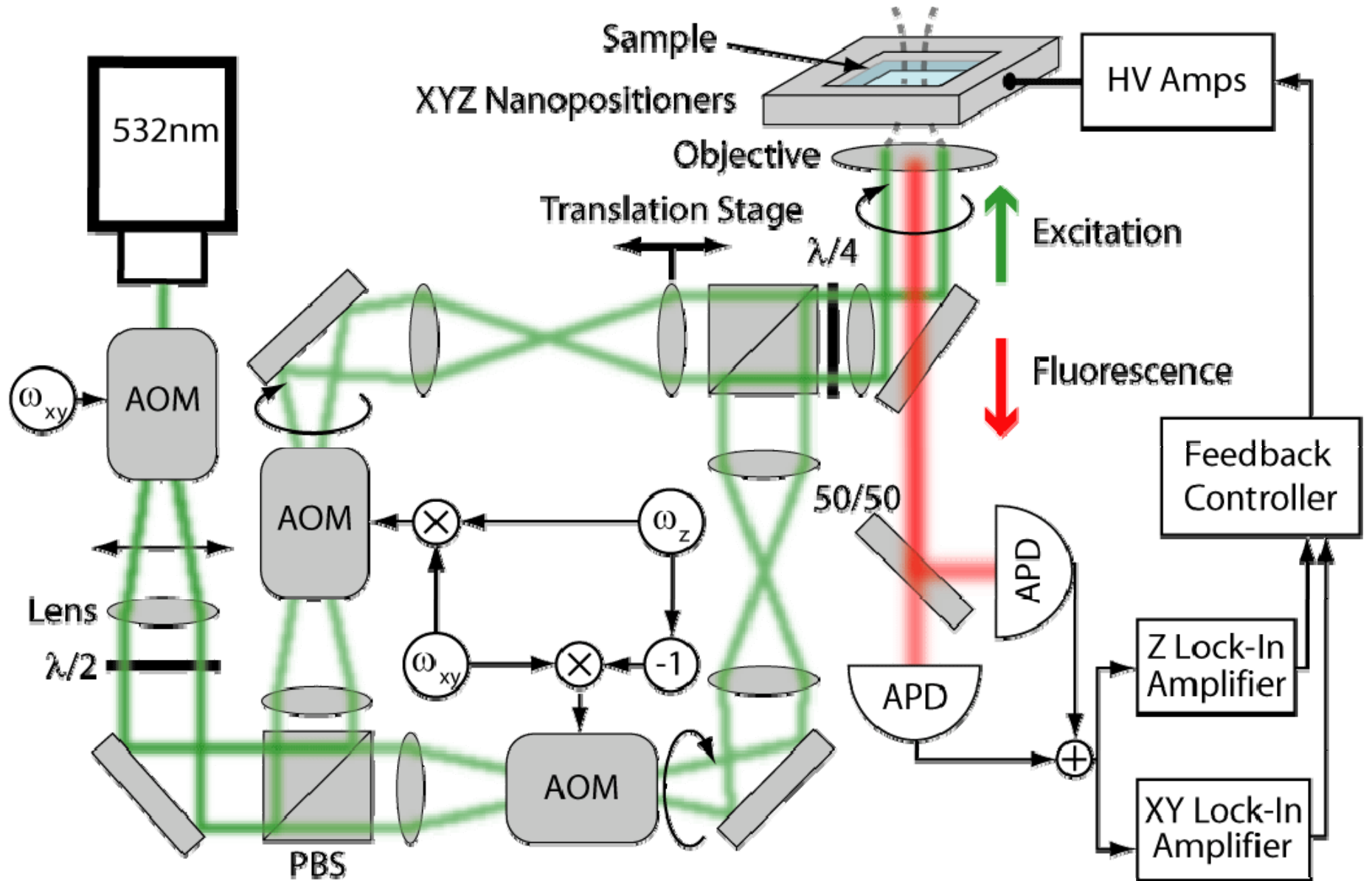


3-D tracking

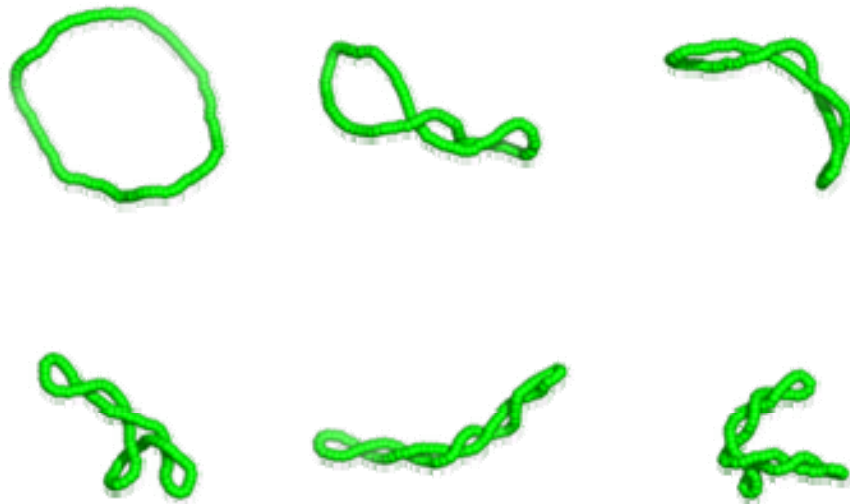


Schematic apparatus

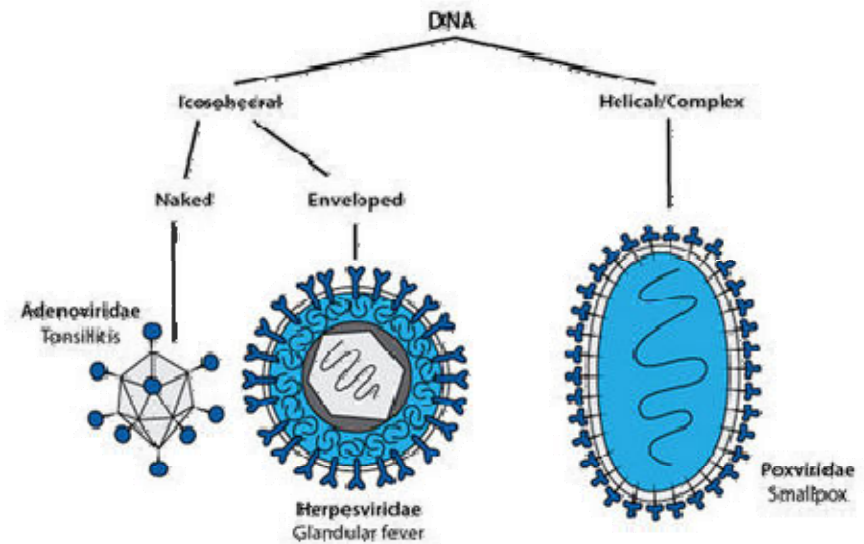
Kevin McHale, A. J. Berglund and HM, Nano Lett. **7**, 3535 (2007)



Free-solution dynamics of large dsDNA molecules



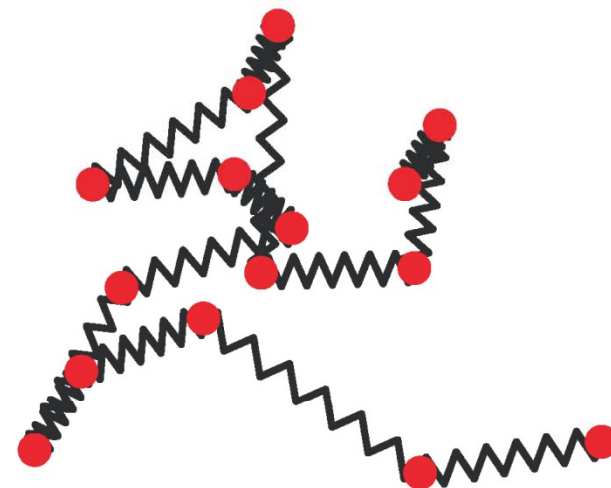
A. J. Spakowitz, Stanford University



www.sciencelearn.org.nz

“Simple” (linearized) polymer models

- ▶ Rouse: Beads and springs
- ▶ Zimm: Hydrodynamic couplings
- ▶ Semi-flexible: Bending energy
- ▶ None: excluded volume



Question: which (if any) model quantitatively describes DNA?

Experiments on genomic lambda-DNA

K. McHale and HM, submitted

References

- ▶ Initial D measurements: Smith, Perkins & Chu, *Macromolecules*, 1995
- ▶ Intramolecular fluorescence on trapped DNA: Quake, Babcock & Chu, *Nature*, 1997
- ▶ Intramolecular FCS of DNA: Lumma, Keller, Vilgis & Rädler, *PRL*, 2003; Shusterman, Alon, Gavrinov & Krichevsky, *PRL*, 2004; Winkler, Keller & Rädler, *PRE*, 2006; Petrov, Ohrt, Winkler & Schwille, *PRL*, 2006
- ▶ DNA in ABEL trap: Cohen & Moerner, *PRL* and *PNAS*, 2007

Lumma *et al.*, Figure 1

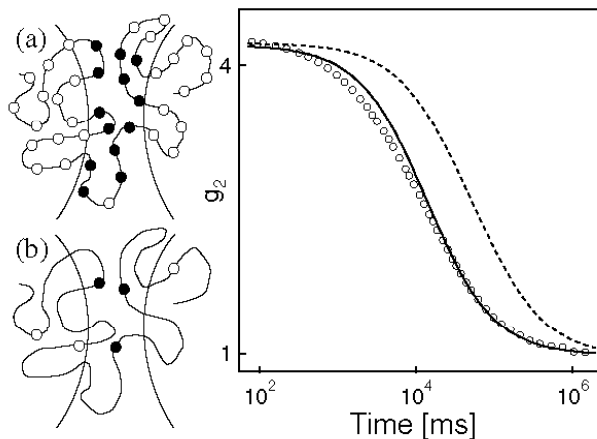
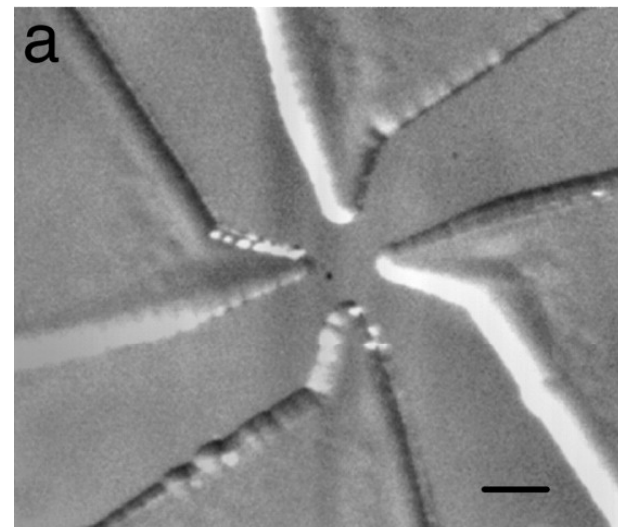


FIG. 1. The measured autocorrelation curve g_2 for dilute λ -phage DNA deviates from the correlation function expected for a stiff finite-size polymer with $R_g = 0.73 \mu\text{m}$ (dashed line). The solid line shows the poor agreement with a best fit to the correlation function of a pointlike particle. The drawings illustrate labeled DNA as a chain of linked chromophores for (a) high and (b) low average label densities.

A. E. Cohen and W.E. Moerner
Proc. Natl. Acad. Sci. U.S.A. **103**, 4362 (2006)



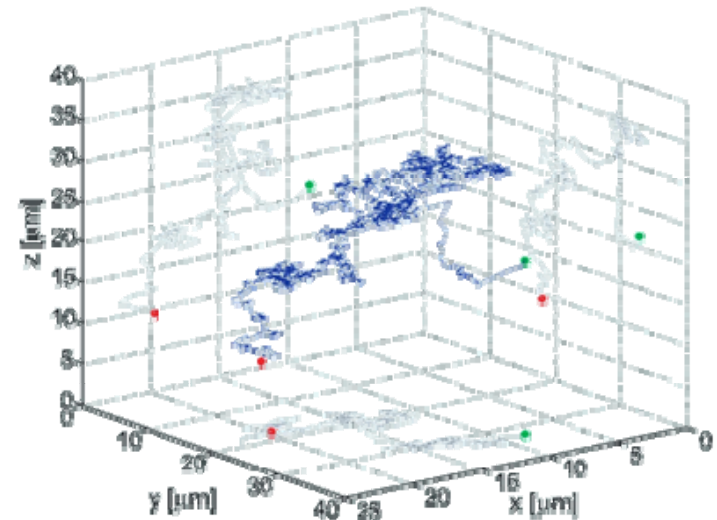
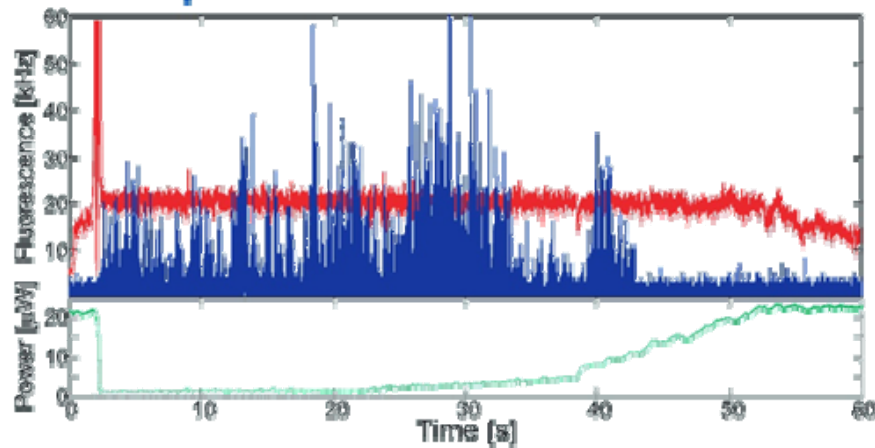
A. E. Cohen and W. E. Moerner, Phys. Rev. Lett. **98**, 116001 (2007)



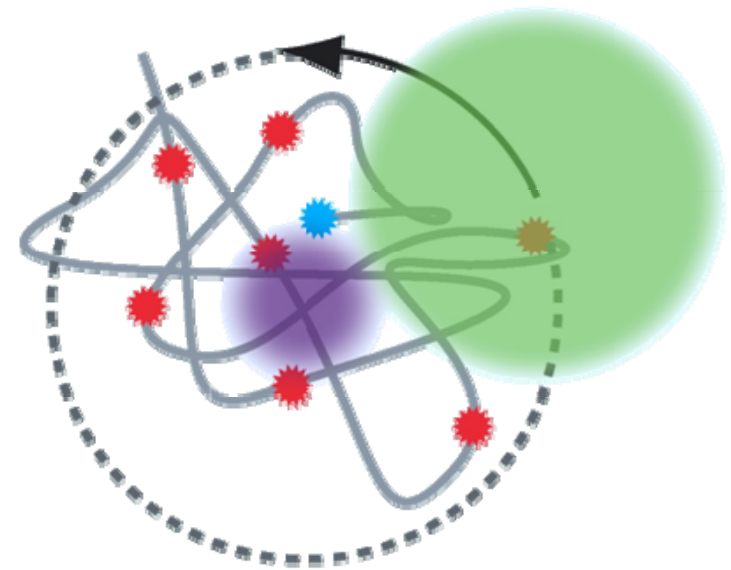
Experiments on genomic lambda-DNA

K. McHale and HM, submitted

Example data

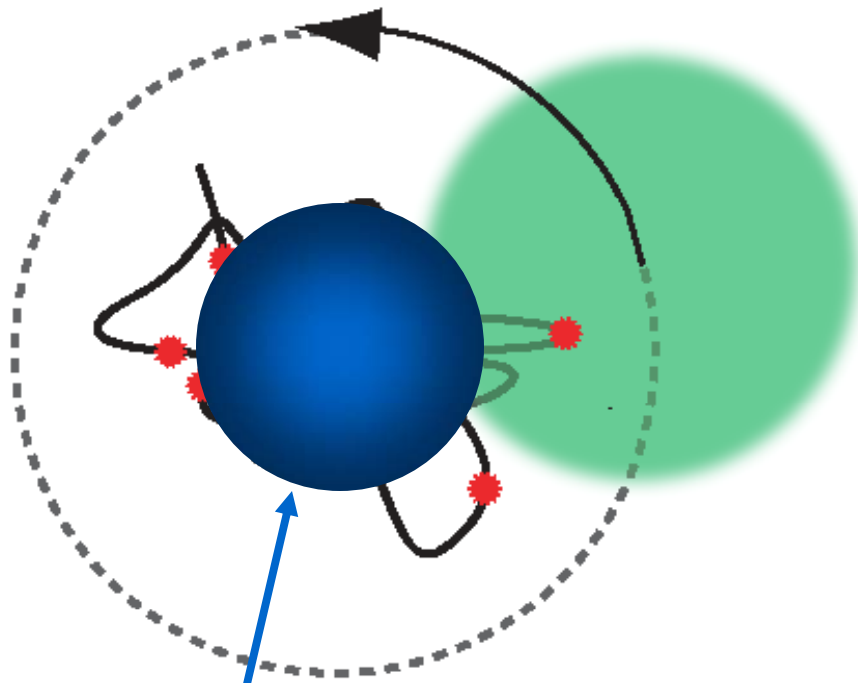


- ▶ Dense tracking labels (POPO-3)
- ▶ Single probe label (Atto425)
- ▶ Fluorescence rate and beam intensity stabilization
- ▶ Alternating excitation to prevent cross-talk

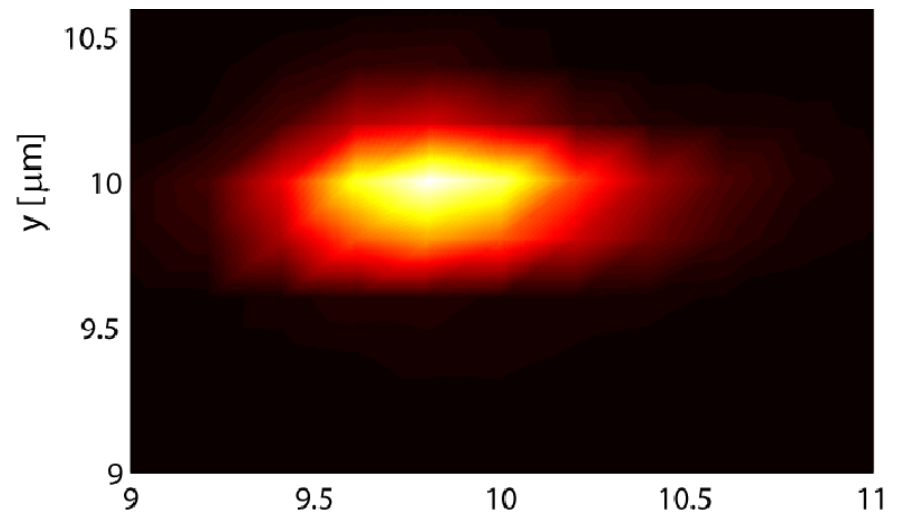
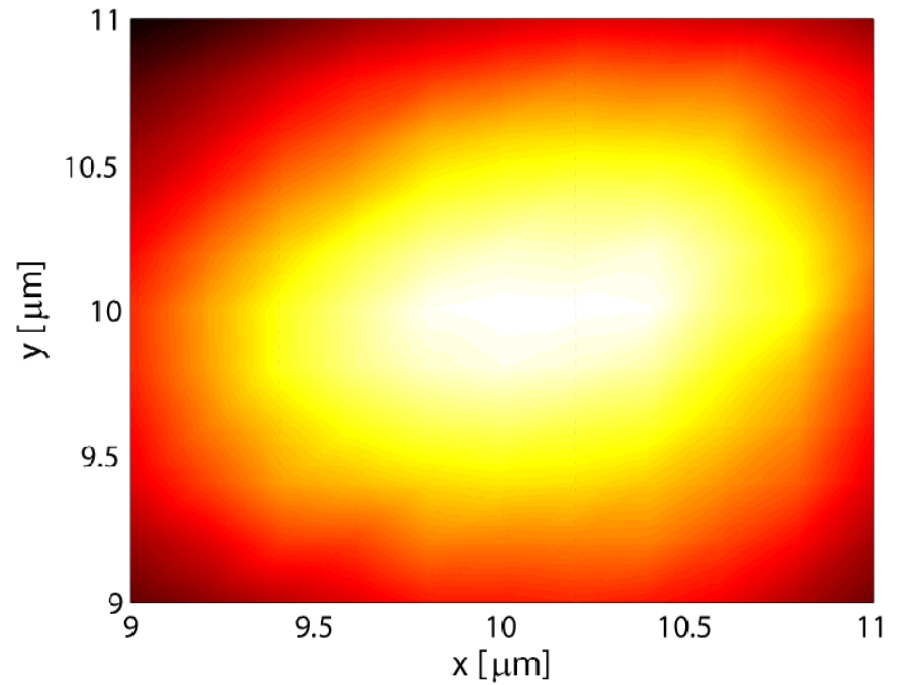


Two-color alternating excitation

K. McHale and HM, submitted



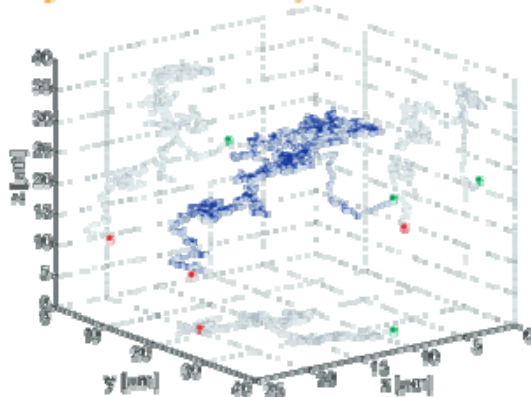
- improves FCS sensitivity
- decouples tracking from FCS
- could be used for FRET/catalysis



Center-of-mass motion

K. McHale and HM, submitted

Get systematic parameters and D from stage motion

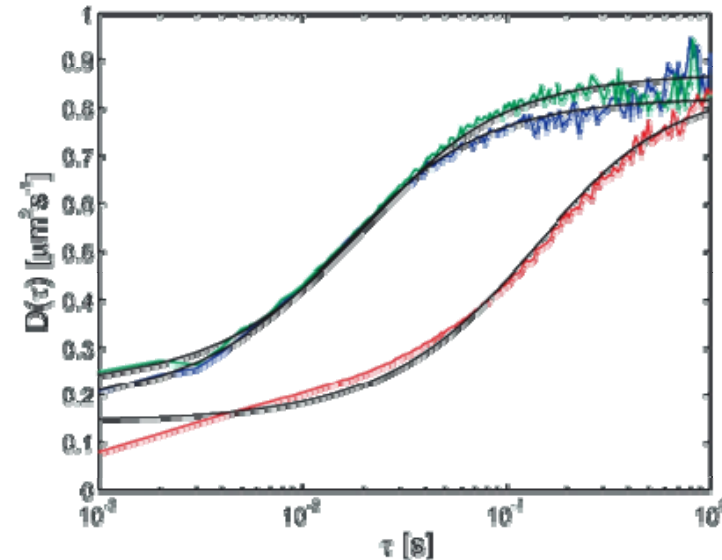


Increment statistics:

$$\hat{D}(\tau) \equiv \frac{1}{2\tau} \left\langle (Y_{t+\tau} - Y_t)^2 \right\rangle$$

- ▶ Tracking bandwidths: $\gamma_{xy} = 15\text{Hz}$, $\gamma_z = 2\text{Hz}$
- ▶ RMS localization: $L_{xy} = 110\text{nm}$, $L_z = 290\text{nm}$
- ▶ Diffusion coefficient: $D = (0.80 \pm 0.05)\mu\text{m}^2/\text{s}$

D inconsistent with all other measurements!
(but really can't be wrong...)



Intramolecular motion

K. McHale and HM, submitted

$$\langle r_{t+\tau} \cdot r_t \rangle = \frac{6r_0^2}{\pi^2} \sum_{q=1}^{\infty} \frac{1}{q^2} e^{-\tau/\tau_q}$$

$$g_x(\tau) = \left[1 - \left(\frac{\sigma^2 e^{-\gamma\tau} + \frac{1}{3} \langle r_{t+\tau} \cdot r_t \rangle}{\sigma^2 + \frac{1}{3} r_0^2 + \frac{1}{4} w^2} \right) \right]^{-1/2}$$

Fit parameters:

- ▶ r_0 : size parameter ($R_g \sqrt{2}$)
- ▶ τ_1 : characteristic time
- ▶ h : *draining* parameter

Interpretation

- ▶ (r_0, τ_1) theoretically consistent with $D \approx 0.8 \mu\text{m}^2/\text{s}$
- ▶ Data and R/Z theories internally consistent to a factor of 2-4
- ▶ **Zimm model is inconsistent with the FCS data!**

