

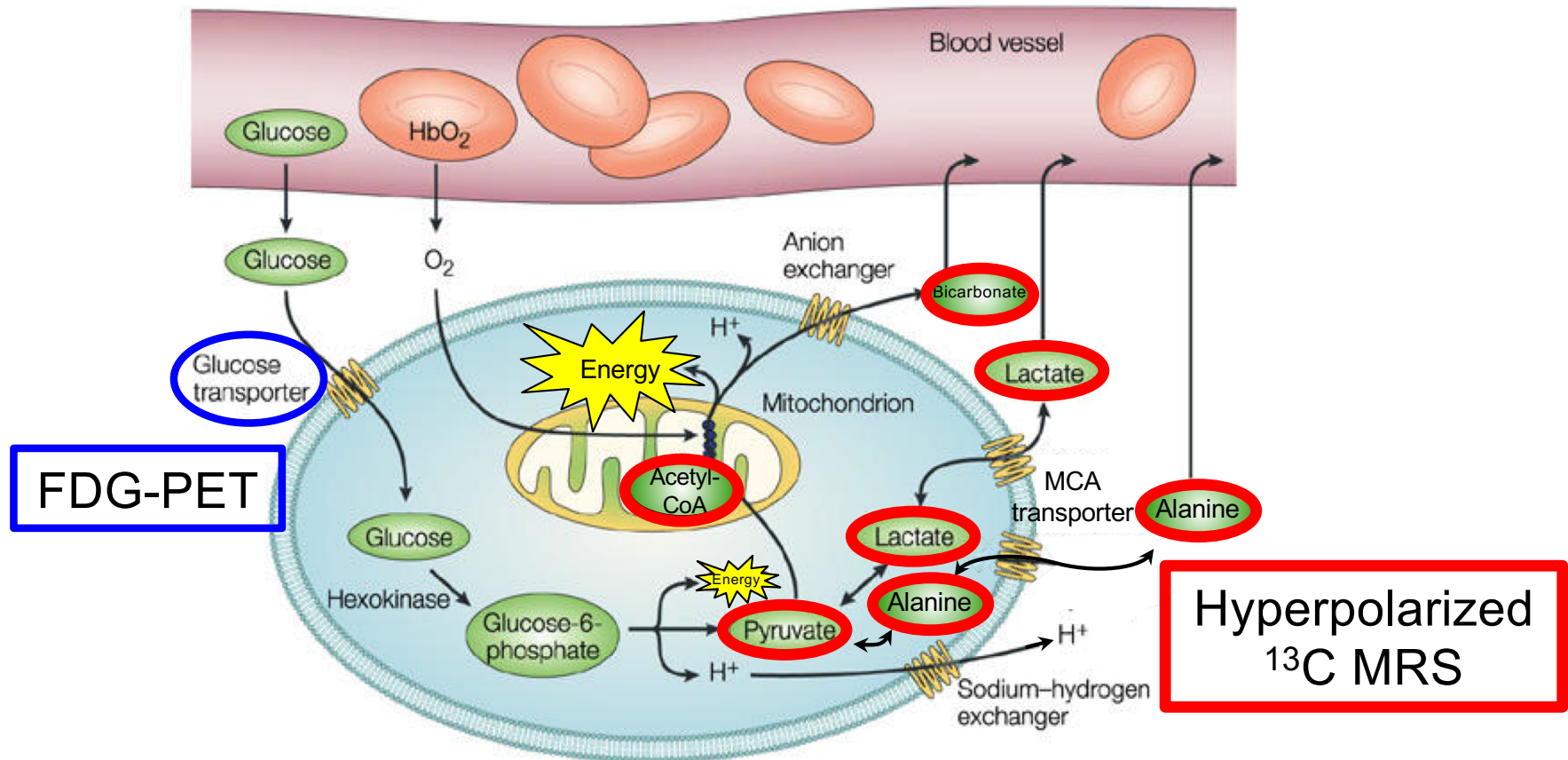
# Lecture #9

## Redfield theory: Examples

- Topics
  - Hyperpolarized  $^{13}\text{C}$ -urea
  - Hyperpolarized  $^{13}\text{C}$ -pyruvate
- Handouts and Reading assignments
  - Shang, et al., “Handheld Electromagnet Carrier for Transfer of Hyperpolarized Carbon-13 Samples”, MRM, early view, 2015.
  - Lau, et al., “A calibration-based approach to real-time in vivo monitoring of pyruvate C1 and C2 polarization using the JCC spectral asymmetry”, NMR Biomed., 2013; 26.

# Hyperpolarized $^{13}\text{C}$ MRS

- Images metabolism by performing an in vivo tissue assay.
- Key idea: inject a biological substrate and image both the substrate and its downstream metabolic products.

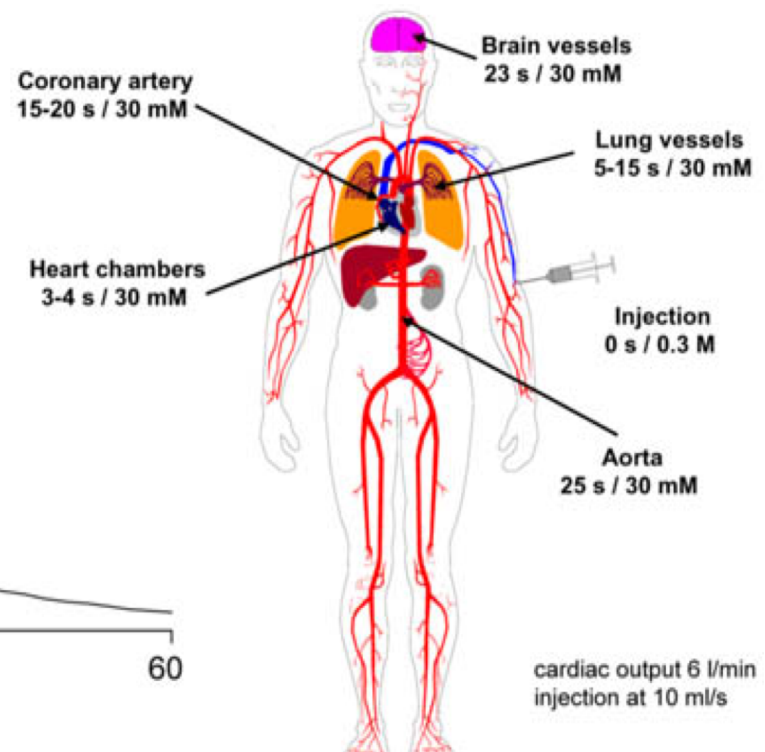
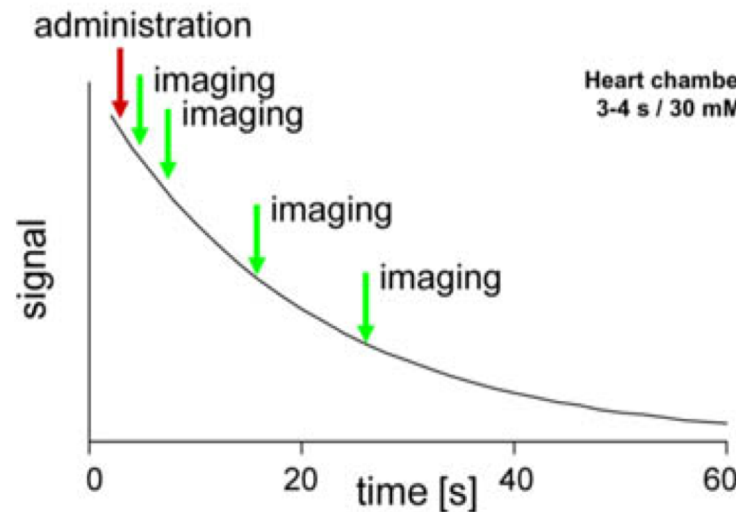
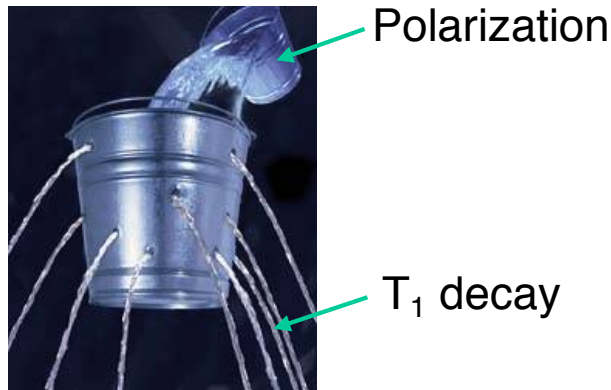


Key technology: A polarizer that magnetically prepares the substrate to boost its MR visibility by >10,000 fold.

# In Vivo Imaging Requirements

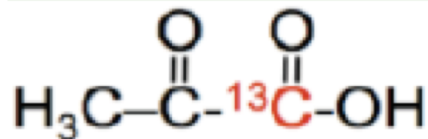
- Low toxicity (mM conc.)
- Long NMR relaxation times
- Chemical shift separation
- Rapid cellular uptake
- Rapid metabolism

Signal decays by relaxation and dilution



➔ Focus on low molecular weight endogenous compounds.

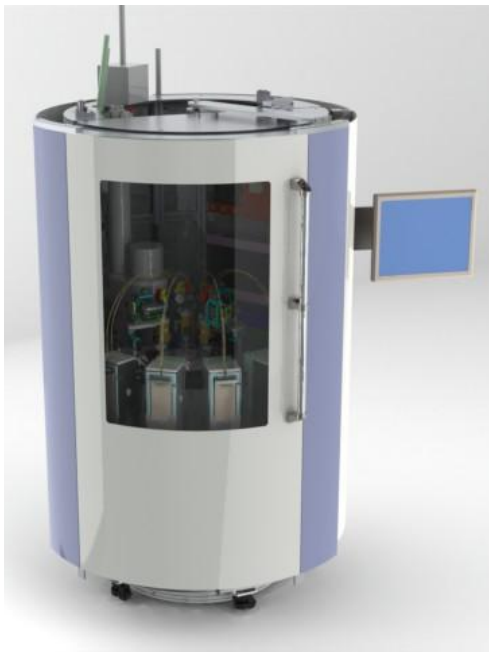
Example: [1- $^{13}\text{C}$ ]pyruvate



25% polarization ~ 30,000 fold signal gain!  
In vivo  $T_1 = 30$  s

# Hyperpolarized Carbon-13 Experiment

Hyperpolarization



1.5 - 3 hours

Run/walk and inject



~30 seconds

Scan



1-2 minutes

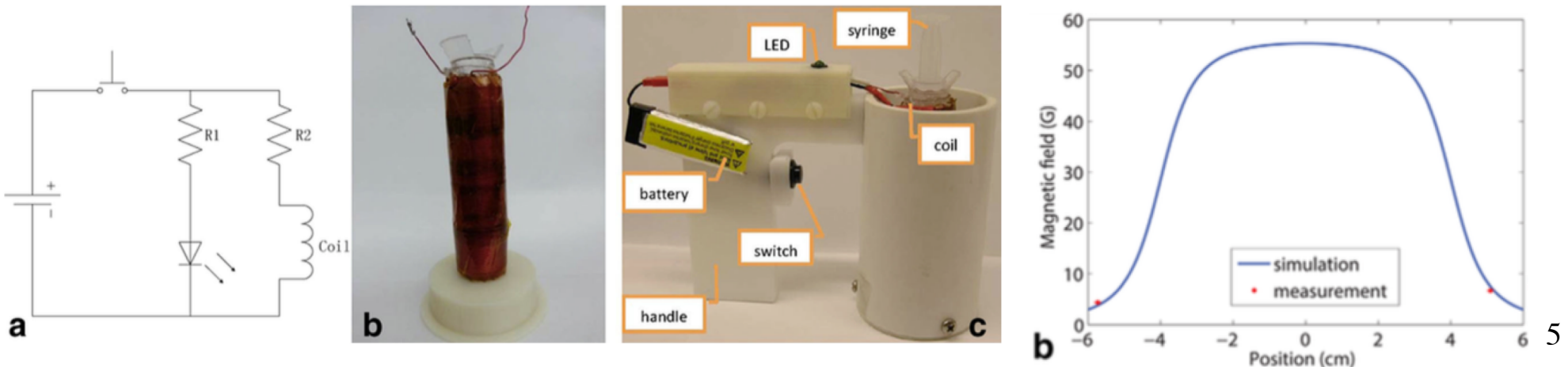
# Example 1

**NOTE**

Magnetic Resonance in Medicine 00:00–00 (2015)

## Handheld Electromagnet Carrier for Transfer of Hyperpolarized Carbon-13 Samples

Hong Shang,<sup>1,2</sup> Timothy Skloss,<sup>3</sup> Cornelius von Morze,<sup>1</sup> Lucas Carvajal,<sup>1</sup>  
Mark Van Criekinge,<sup>1</sup> Eugene Milshteyn,<sup>1,2</sup> Peder E. Z. Larson,<sup>1,2</sup>  
Ralph E. Hurd,<sup>3</sup> and Daniel B. Vigneron<sup>1,2\*</sup>

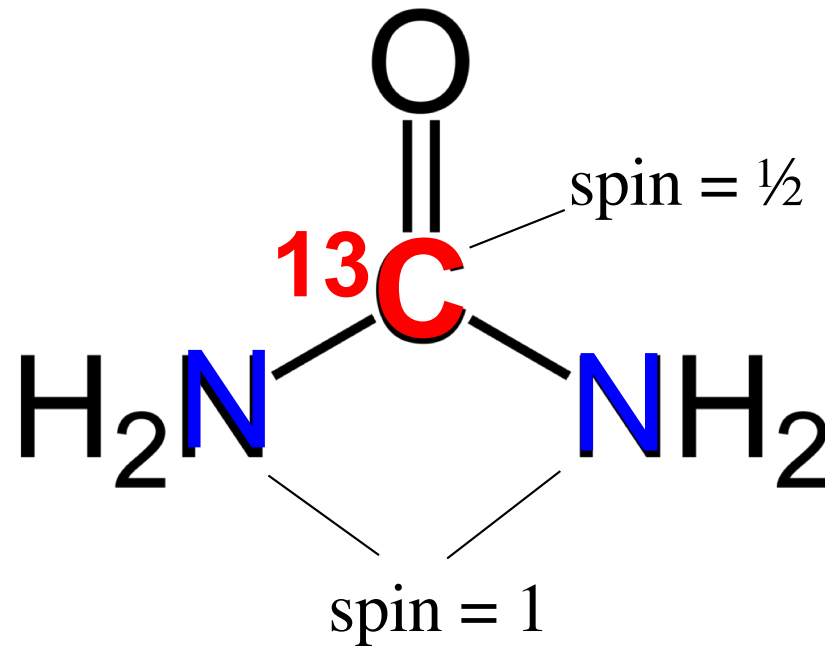


# Hyperpolarized $^{13}\text{C}$ MRS

- “Some HP  $^{13}\text{C}$  substrates can lose polarization extremely quickly in low magnetic field when they are transferred between the polarizer and the MR scanner, reducing the SNR.” – Shang, et al.
- Which substrates?
- Why is this a “low magnetic field” effect?
- How low is “low”?

# [<sup>13</sup>C]urea

- Hyperpolarized [<sup>13</sup>C]urea of interest for measuring perfusion



- Scalar coupling between fast-relaxing spin 1 quadrupolar-coupled <sup>14</sup>N and spin 1/2 <sup>13</sup>C nuclei results in rapid loss of polarization at low field.

# Scalar relaxation of the 2<sup>nd</sup> kind

- From the homework, we derived: 
$$\frac{1}{T_{1,SC2}} = \frac{2(2\pi J)^2 S(S+1)}{3} \frac{T_{2,S}}{1 + (\omega_I - \omega_s)^2 T_{2,S}^2}$$

- For hyperpolarized [<sup>13</sup>C-<sup>14</sup>N<sub>2</sub>]urea:

Extra factor of 2 due to two <sup>14</sup>N nuclei.

Compare to Shang, et al. Eqn [1].

$$\frac{1}{T_1} = \frac{1}{T_{1,0}} + \frac{2 \cdot 8\pi^2 J^2 S(S+1)}{3} \frac{T_{2,N}}{1 + (\gamma_C B_0 - \gamma_N B_0)^2 T_{2,N}^2} = \frac{1}{T_{1,0}} + \frac{32\pi^2 J^2}{3} \frac{T_{2,N}}{1 + B_0^2 (\gamma_C - \gamma_N)^2 T_{2,N}^2}$$

Relaxation from other mechanisms

- Some numbers...

$$S = 1$$

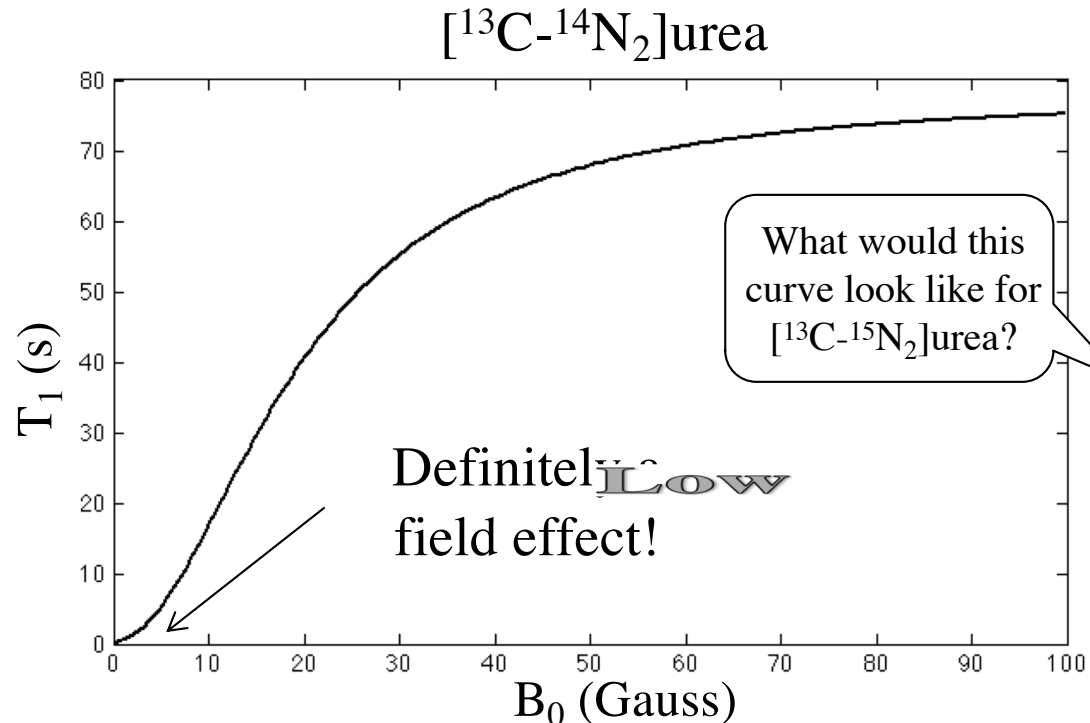
$$J = 14.5 \text{ Hz}$$

$$T_{1,0} = 78 \text{ s}$$

$$T_{2,N} = 2 \times 10^{-4} \text{ s}$$

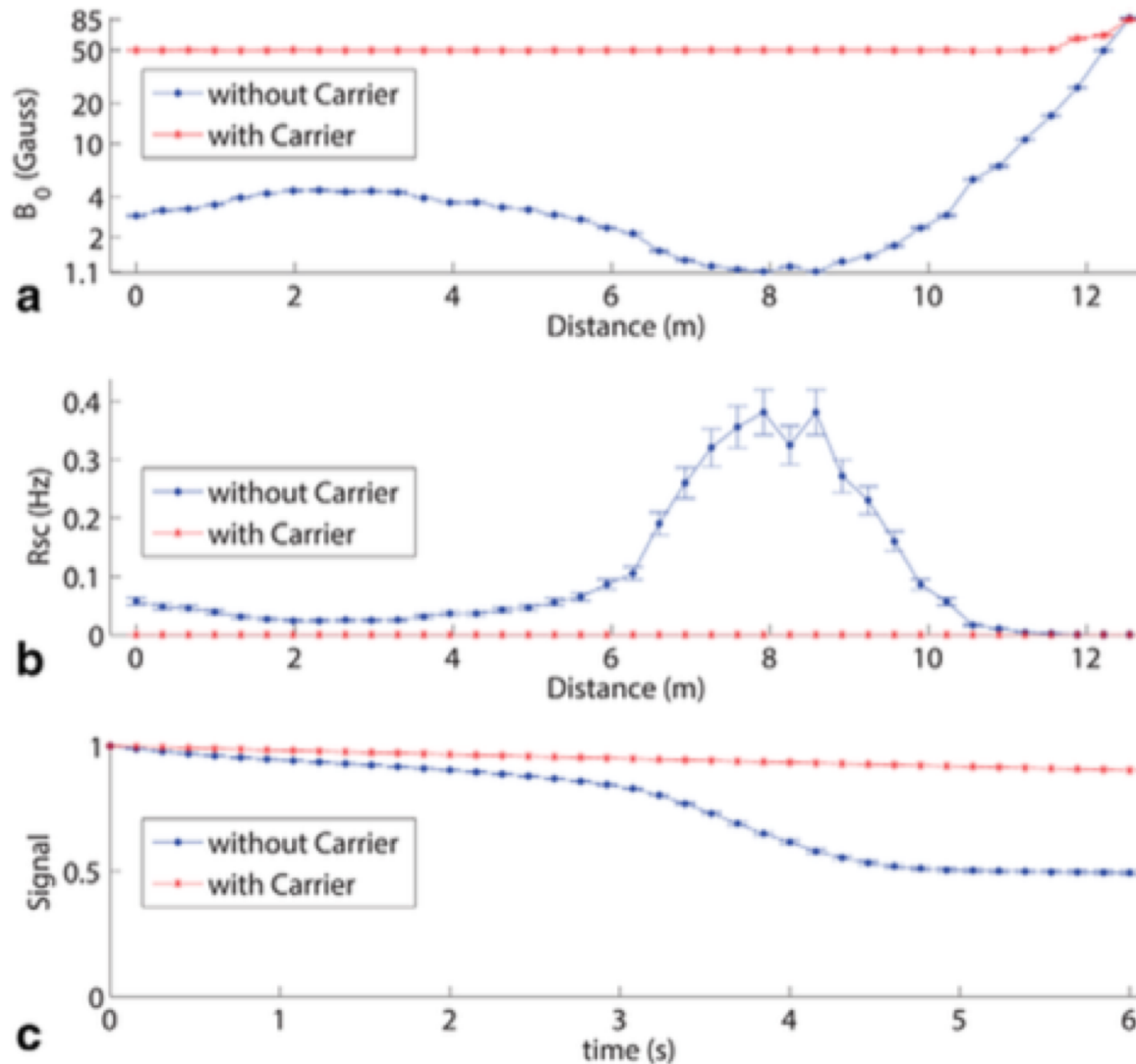
$$\gamma_N = 19.331 \times 10^6 \text{ rad/s/Tesla}$$

$$\gamma_C = 67.262 \times 10^6 \text{ rad/s/Tesla}$$

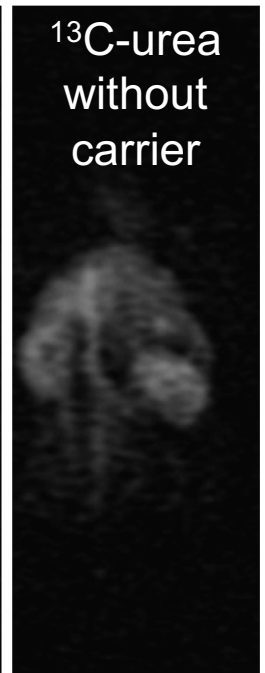
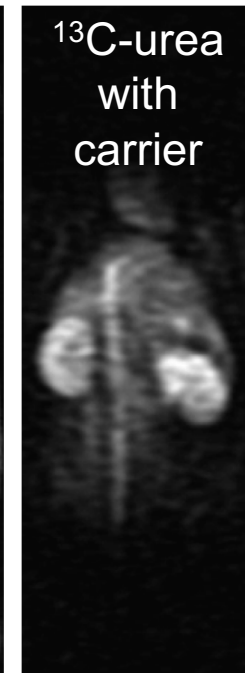
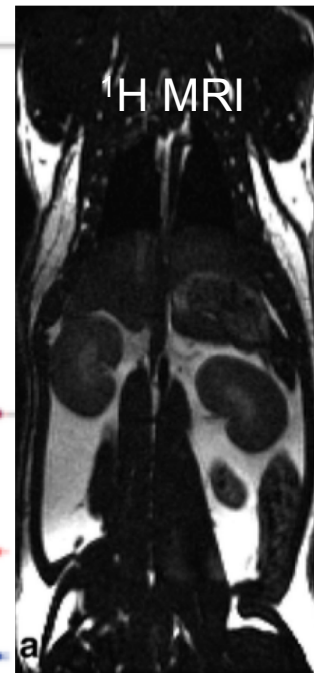




# Hyperpolarized $^{13}\text{C}$ -urea

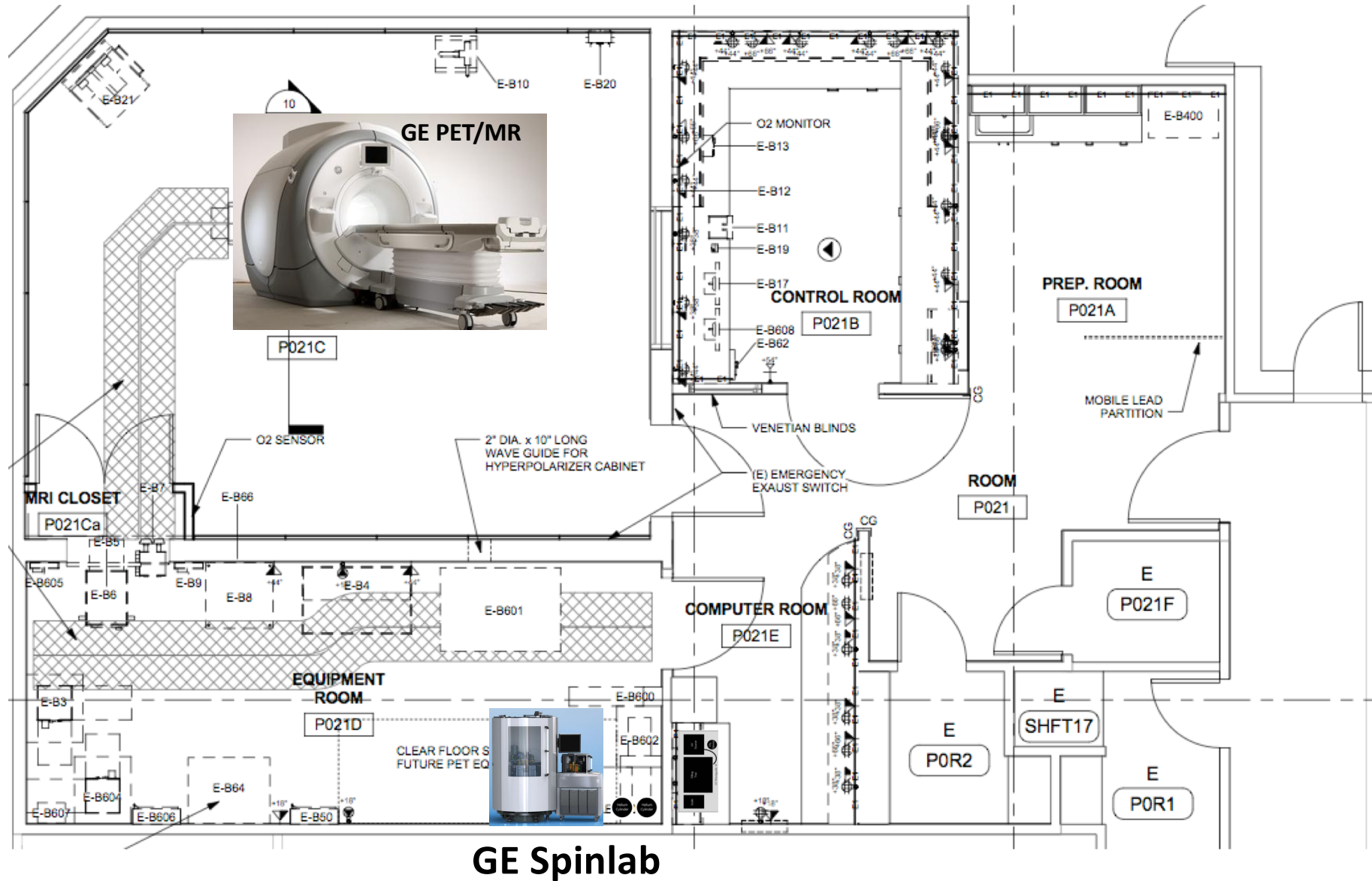


Rat



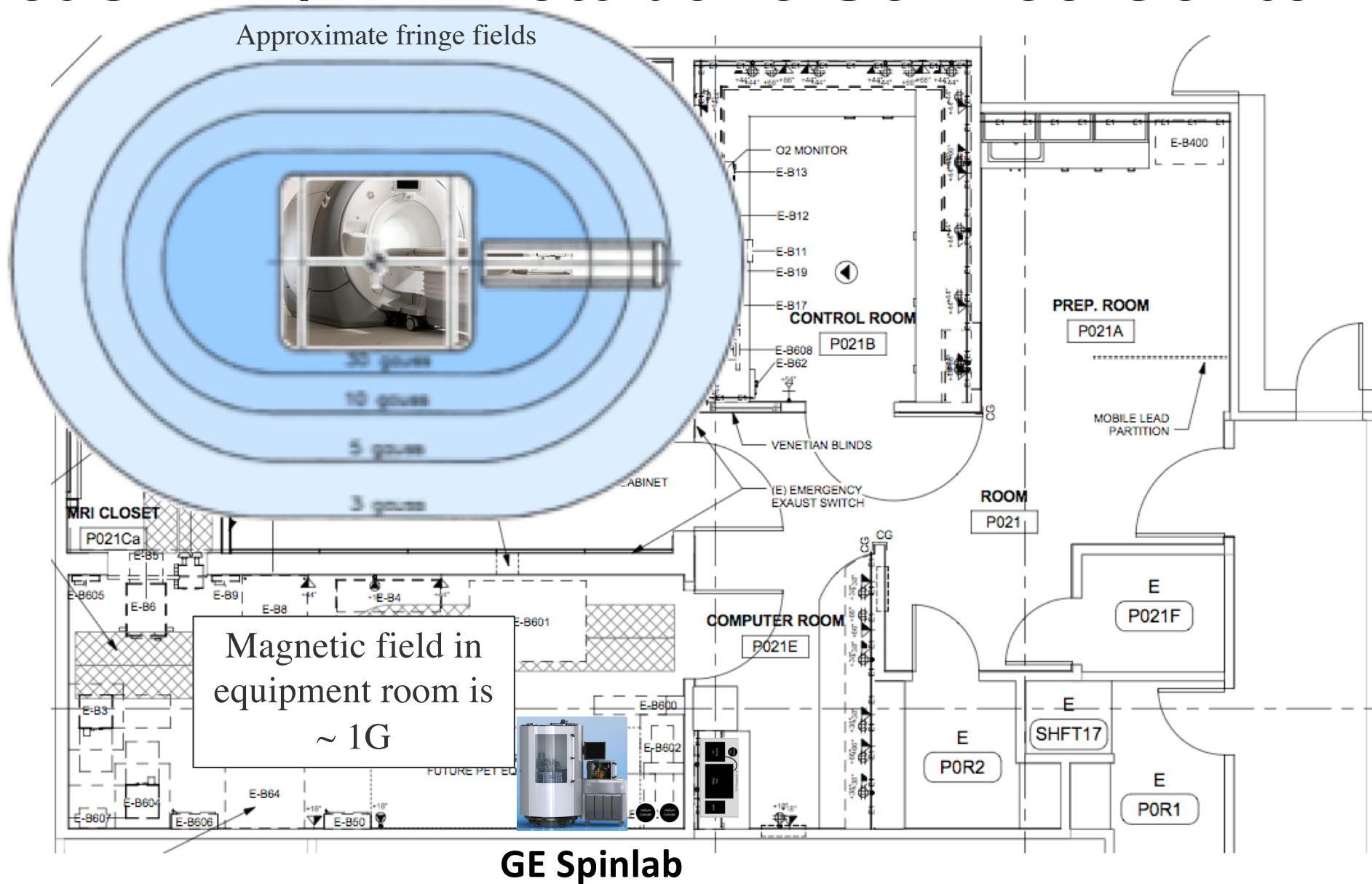
Shang, et al., "Handheld Electromagnet Carrier for Transfer of Hyperpolarized Carbon-13 Samples", MRM, early view, 2015

# Lucas PET/MR Metabolic Service Center



PET/MR + Hyperpolarized MRS

# Lucas PET/MR Metabolic Service Center



PET/MR + Hyperpolarized MRS

# Ex 2: Hyperpolarized [1,2-<sup>13</sup>C]Pyr

**NMR**  
IN BIOMEDICINE

Research article

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(wileyonlinelibrary.com) DOI: 10.1002/nbm.2942

## A calibration-based approach to real-time *in vivo* monitoring of pyruvate C<sub>1</sub> and C<sub>2</sub> polarization using the $J_{CC}$ spectral asymmetry

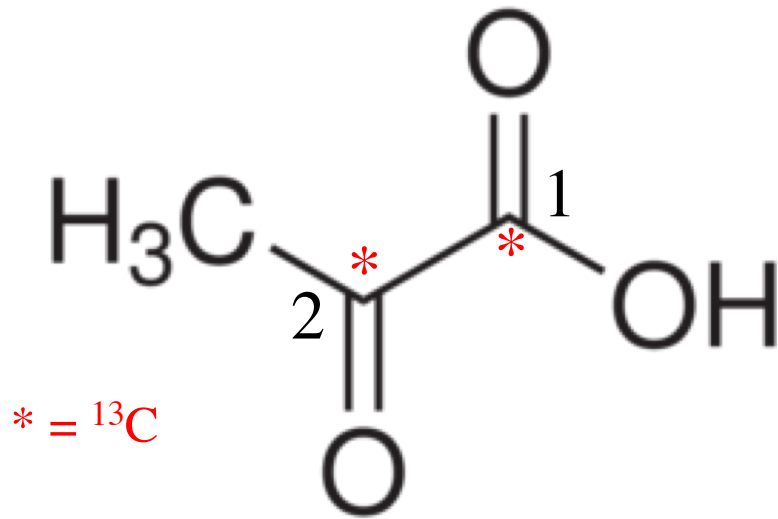
Justin Y. C. Lau<sup>a,b</sup>, Albert P. Chen<sup>c</sup>, Yi-Ping Gu<sup>b</sup> and Charles H. Cunningham<sup>a,b,\*</sup>

A calibration-based technique for real-time measurement of pyruvate polarization by partial integral analysis of the doublet from the neighbouring  $J$ -coupled carbon is presented. *In vitro* calibration data relating the C<sub>2</sub> and C<sub>1</sub> asymmetries to the instantaneous C<sub>1</sub> and C<sub>2</sub> polarizations, respectively, were acquired in blood. The feasibility of using the *in vitro* calibration data to determine the instantaneous *in vivo* C<sub>1</sub> and C<sub>2</sub> polarizations was demonstrated in the analysis of rat kidney and pig heart spectral data. An approach for incorporating this technique into *in vivo* protocols is proposed. Copyright © 2013 John Wiley & Sons, Ltd.

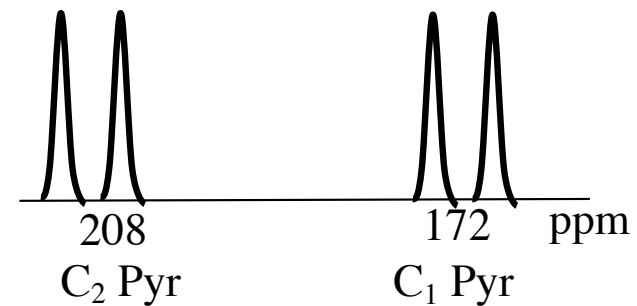
**Keywords:** hyperpolarized; pyruvate; DNP; carbon-13; polarization measurement; asymmetry

# [1-<sup>13</sup>C]Pyruvate

- 1% of [1-<sup>13</sup>C]Pyr is actually doubly labeled [1,2-<sup>13</sup>C]Pyr due to the natural abundance of <sup>13</sup>C.



<sup>13</sup>C MRS spectrum



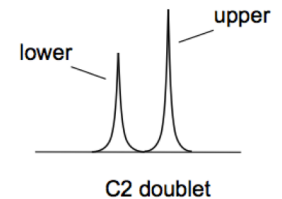
- Carbon-carbon J-coupling of [1,2-<sup>13</sup>C]Pyr leads to doublet resonances
- At standard temperatures, doublets are symmetric



Which carbon will have the longest T<sub>1</sub>?

# Doublet Asymmetry

- Hamiltonian:  $\hat{H}_0 = -\omega_I \hat{I}_z - \omega_S \hat{S}_z + 2\pi J (\hat{I} \cdot \hat{S})$
- Spin density operator:  $\hat{\sigma}_0 \approx \frac{1}{4} \hat{E} + \frac{1}{2} P_C \hat{I}_z + \frac{1}{2} P_C \hat{S}_z + \frac{1}{2} P_C^2 2\hat{I}_z \hat{S}_z$   $P_C = \text{carbon polarization}$
- Consider an experiment where the initial carbon polarization is  $P_C$  and the flip angle for the  $C_1$ - and  $C_2$ -carbons of  $[1,2-^{13}\text{C}]\text{Pyr}$  is  $\phi$ .
- The  $C_2$  doublet (as well as the  $C_1$  doublet) is asymmetric primarily due to the  $2\hat{I}_z \hat{S}_z$  term generating an anti-phase signal (with a minor contribution from residual strong coupling effects).
- Defining an asymmetry parameter  $a_c = (\text{upper peak} - \text{lower peak}) / (\text{upper peak} + \text{lower peak})$ , one can show that at  $t = 0$ ,  $a_c$  is given by:



$$a_c(0) \approx P_C \cos \phi + \sin \theta$$

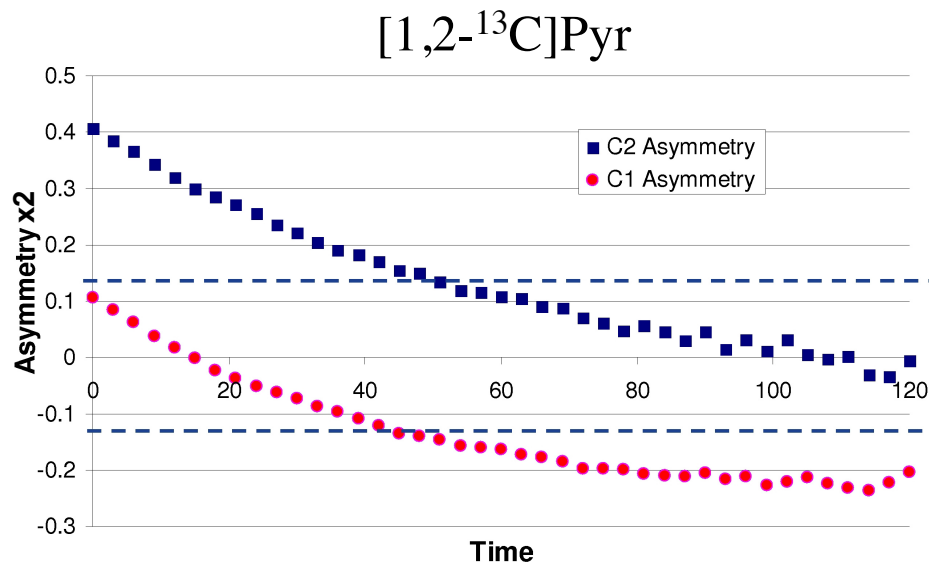
Residual strong coupling parameter = 0.056  
for  $[1,2-^{13}\text{C}]\text{Pyr}$  at 3 T.

# Doublet Asymmetry

- Given that  $P_C$  decays with the  $T_1$  of  $C_2$  pyruvate peak ( $\sim 50$  s in vitro at 3T ), one would expect  $a_C$  to exponentially decay with time constant  $T_1$  towards a thermal equilibrium value of 0.056.

$$a_C(0) \approx P_C \cos \phi + \sin \theta \quad \rightarrow \quad a_C(t) \approx P_C e^{-t/T_1} \cos \phi + \sin \theta?$$

- However, we observe something quite different!



$$a_C(t) \neq P_C e^{-t/T_1} \cos \phi + \sin \theta$$

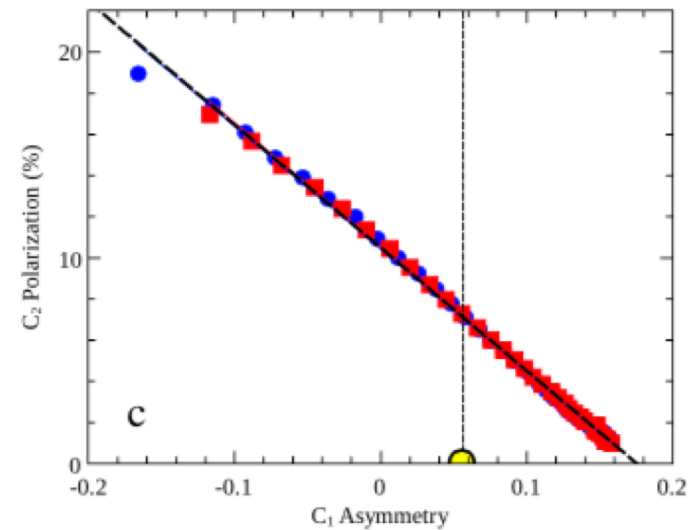
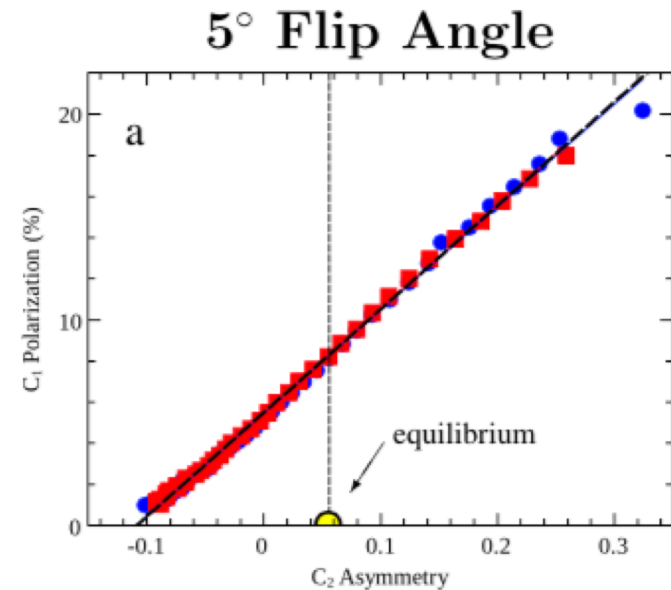
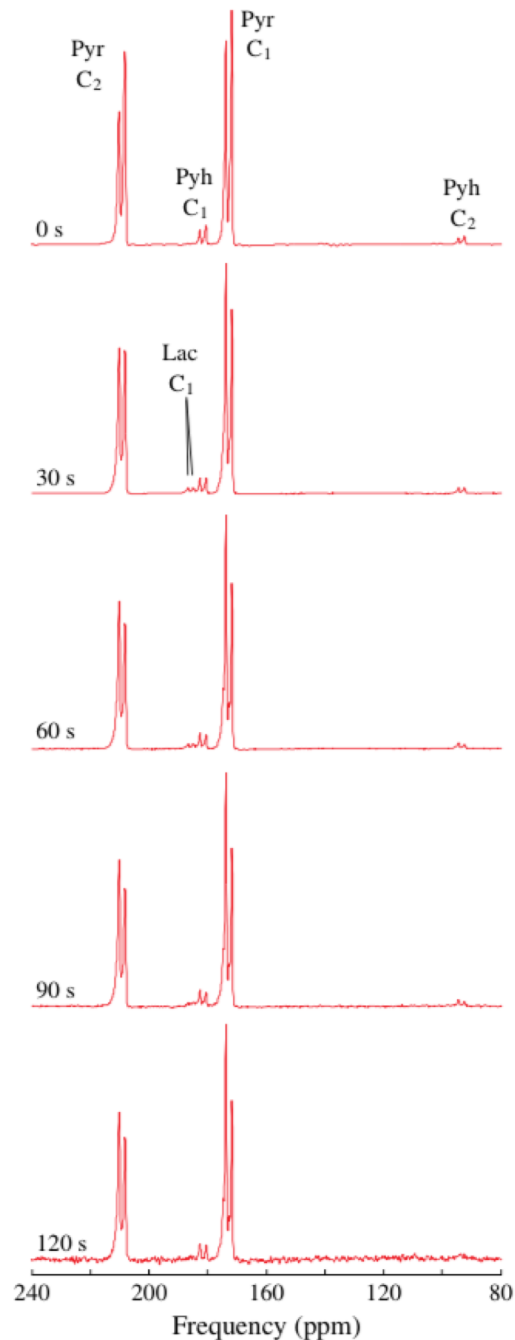
Thermal equilibrium values



Hurd RE, Chen A, et al., Scalar Coupling Patterns In Hyperpolarized Spin Systems: JCC Spectral Pattern In Hyperpolarized 1,2-[ $^{13}C$ ]-Pyruvate, ENC, 2009.

# Hyperpolarized [1,2-<sup>13</sup>C]Pyr

Lau, et al.:

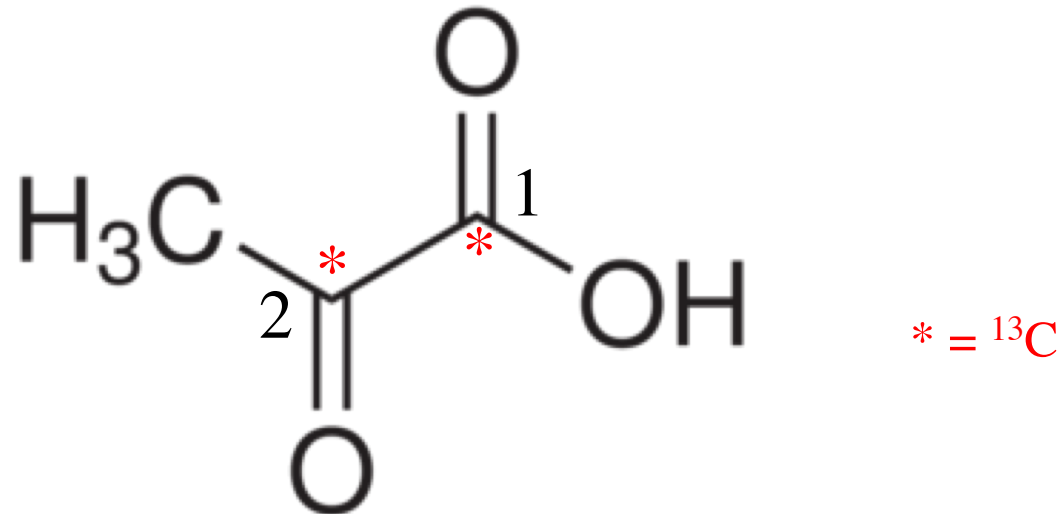


Data doesn't fit proposed analytic model.

$$a_c(t) \neq P_C e^{-t/T_1} \cos \phi + \sin \theta$$



# [1,2-<sup>13</sup>C]Pyruvate



- If we write  $a_C(t)$  in terms of the coherences, we get...

$$a_{C_2}(t) = \frac{\overline{\langle 2\hat{I}_z\hat{S}_z \rangle}(t)}{\overline{\langle \hat{S}_z \rangle}(t)} \cos\phi + \frac{1}{2} \frac{\left( \overline{\langle \hat{I}_z \rangle}(t) + \overline{\langle \hat{S}_z \rangle}(t) \right)}{\overline{\langle \hat{S}_z \rangle}(t)} \sin\theta.$$

- What are the possible relaxation mechanisms?

# Time Evolution of Doublet Asymmetry

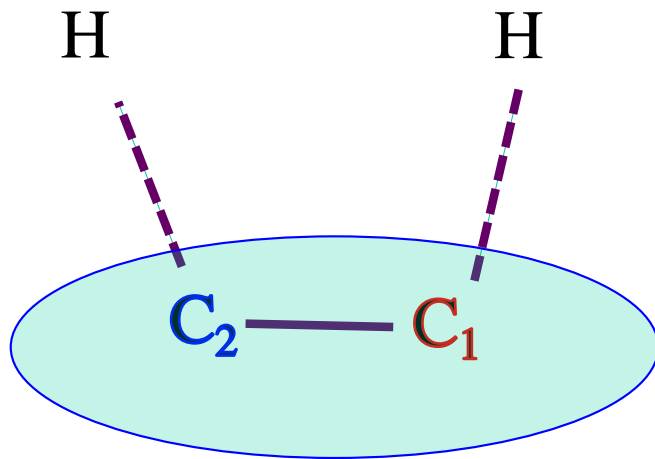
- Relaxation mechanisms to consider:
  1. C<sub>1</sub>-C<sub>2</sub> dipolar
  2. C<sub>1</sub>-H, C<sub>2</sub>-H dipolar
  3. CSA of C<sub>1</sub> and C<sub>2</sub>
  4. Combination of all of the above
- In general...

$$\frac{d}{dt} \begin{bmatrix} \overline{\langle \hat{I}_z \rangle}(t) \\ \overline{\langle \hat{S}_z \rangle}(t) \\ \overline{\langle 2\hat{I}_z \hat{S}_z \rangle}(t) \end{bmatrix} = - \underbrace{\begin{bmatrix} R_{11} & R_{12} & R_{13} \\ R_{21} & R_{22} & R_{23} \\ R_{31} & R_{32} & R_{33} \end{bmatrix}}_{\text{RELAXATION MATRIX}} \begin{bmatrix} \overline{\langle \hat{I}_z \rangle}(t) - I_z^{eq} \\ \overline{\langle \hat{S}_z \rangle}(t) - S_z^{eq} \\ \overline{\langle 2\hat{I}_z \hat{S}_z \rangle}(t) - 2I_z S_z^{eq} \end{bmatrix}$$

- Let's use Redfield relaxation theory to find analytic expressions for the  $R_{ij}$ s.

Note we'll use literature values for any unknown parameters such as  $\tau_C=3\text{ps}$ ,  
 $r=1.1\text{\AA}$ ,  $T_{1,\text{DDCH}}=70\text{s}$ ,  $T_{1,\text{CSA}}=200\text{s}$ .

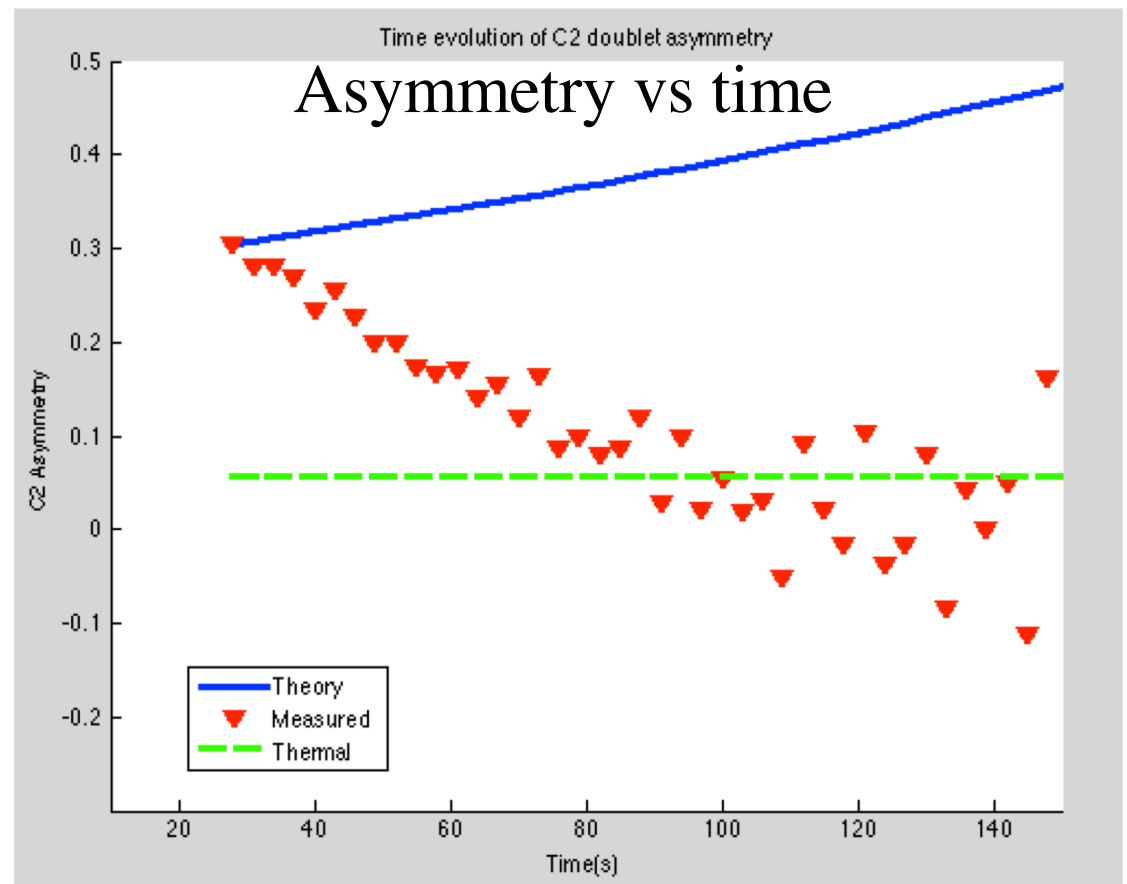
# C<sub>1</sub>-C<sub>2</sub> Dipolar Coupling



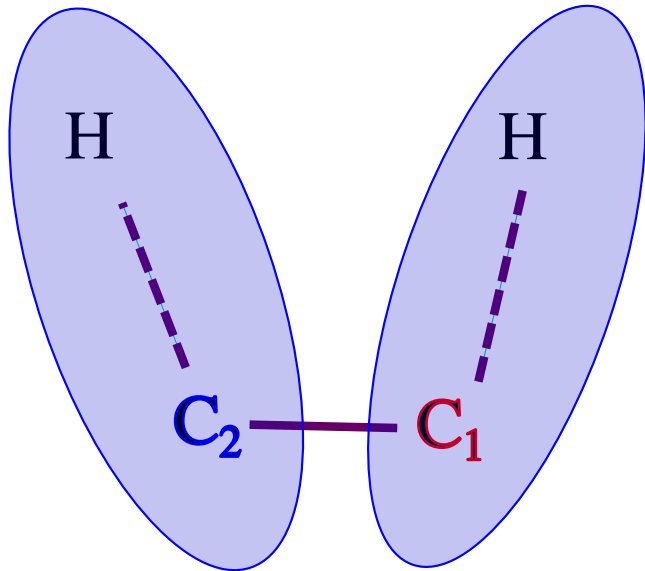
- Not a dominant source of relaxation
- Experimental T<sub>1</sub> values of C<sub>1</sub> in [1,2-<sup>13</sup>C]Pyr similar to [1-<sup>13</sup>C]Pyr

$$\begin{bmatrix}
 \mathbf{R}_{11} & \mathbf{R}_{12} & R_{13} \\
 \mathbf{R}_{21} & \mathbf{R}_{22} & R_{23} \\
 R_{31} & R_{32} & \mathbf{R}_{33}
 \end{bmatrix}$$

RELAXATION MATRIX



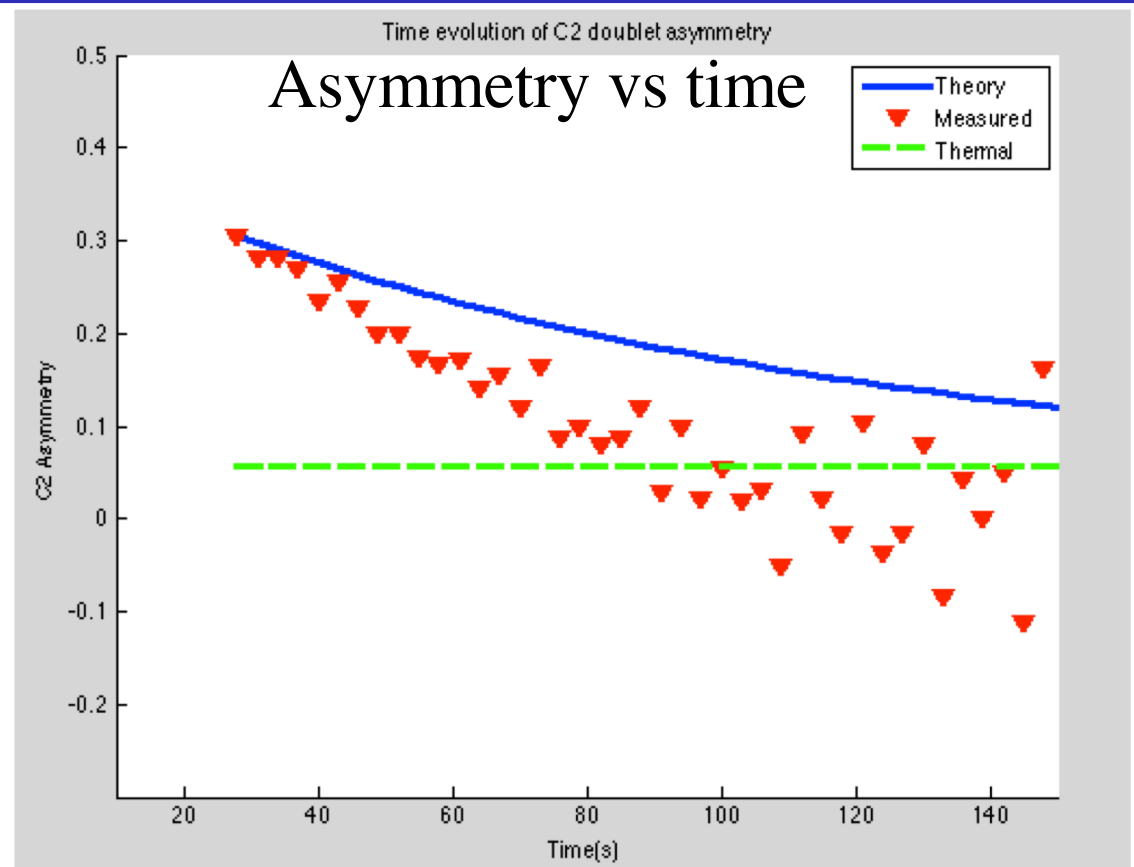
# add C-H Dipolar Coupling



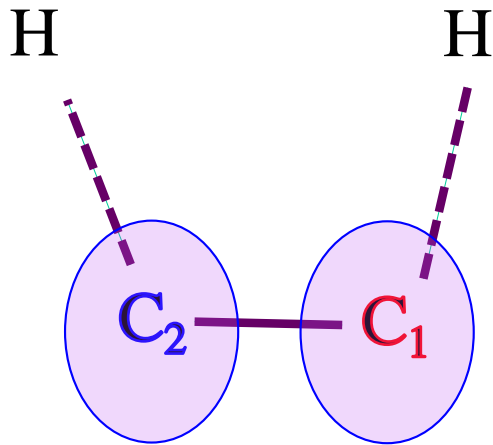
- A major source of relaxation for Pyr
- $^1\text{H}$  is a stronger magnet than  $^{13}\text{C}$
- $2I_Z S_Z$  coherence decays at twice the rate of  $I_Z, S_Z$  coherences
- No field dependence for small molecules

$$\begin{bmatrix} R_{11} & R_{12} & R_{13} \\ R_{21} & R_{22} & R_{23} \\ R_{31} & R_{32} & R_{33} \end{bmatrix}$$

RELAXATION MATRIX



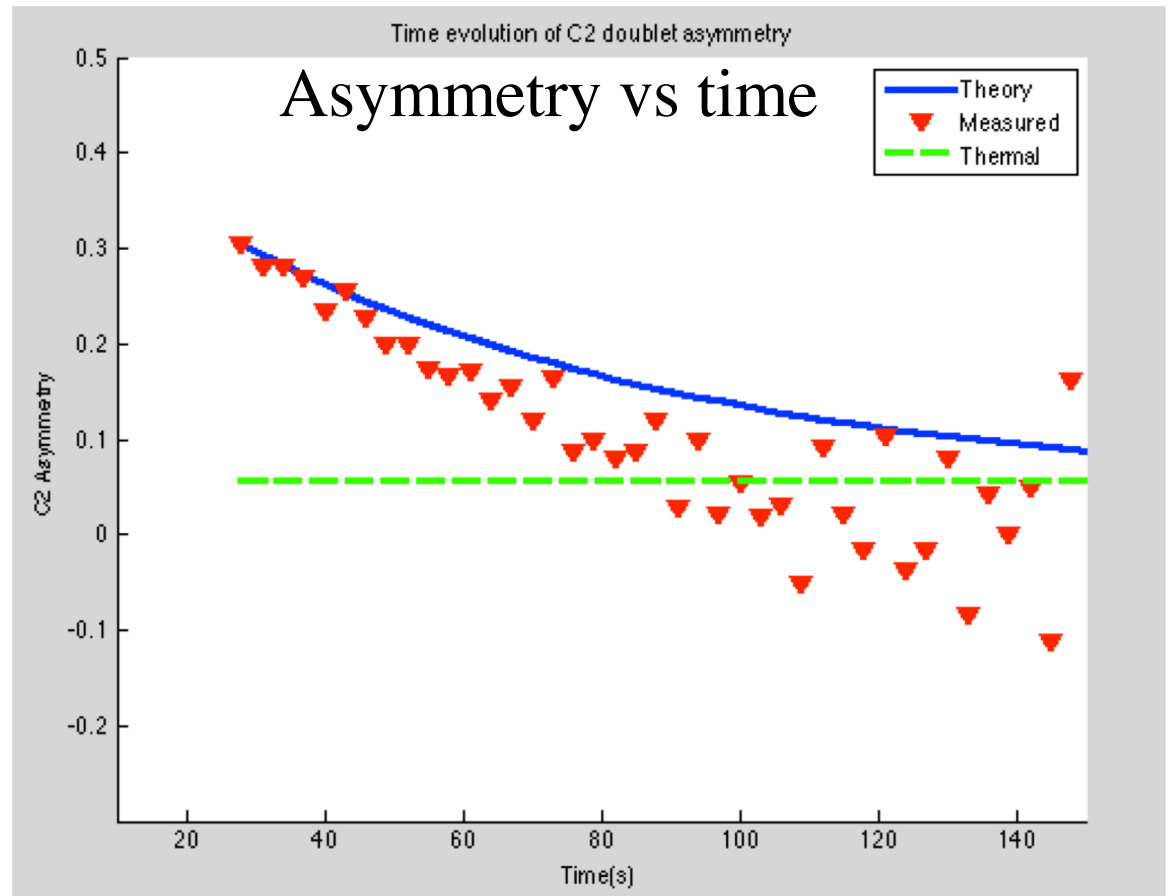
# add Chemical Shift Anisotropy



- A moderate effect at 3T
- Effect is field dependent: C<sub>1,2</sub> T<sub>1</sub> values of [1,2-<sup>13</sup>C]Pyr both decrease with ↑B<sub>0</sub>

$$\begin{bmatrix} R_{11} & R_{12} & R_{13} \\ R_{21} & R_{22} & R_{23} \\ R_{31} & R_{32} & R_{33} \end{bmatrix}$$

RELAXATION MATRIX



# Interference Effects

- When we consider the various relaxation mechanisms occurring simultaneously, some new terms can emerge...
- Consider the case of X = protons dipolar coupled to C<sub>1</sub> and Y=protons dipolar coupled to C<sub>2</sub>

$$\hat{H}_{D_{IX}} = -\frac{\gamma_C \gamma_H \hbar}{r_{IX}^3} \sum_{q=-2}^2 F_q \hat{A}_q \quad \hat{H}_{D_{SY}} = -\frac{\gamma_C \gamma_H \hbar}{r_{SY}^3} \sum_{q=-2}^2 F_q \hat{B}_q$$

$$\begin{aligned} \hat{A}_0 &= \sqrt{\frac{1}{6}} \left( 2\hat{I}_z \hat{X}_z - \frac{1}{2}\hat{I}_+ \hat{X}_- - \frac{1}{2}\hat{I}_- \hat{X}_+ \right) & \hat{B}_0 &= \sqrt{\frac{1}{6}} \left( 2\hat{S}_z \hat{Y}_z - \frac{1}{2}\hat{S}_+ \hat{Y}_- - \frac{1}{2}\hat{S}_- \hat{Y}_+ \right) & F_0 &= \sqrt{\frac{3}{2}} (3\cos^2 \theta - 1) \\ \hat{A}_{\pm 1} &= \frac{1}{2} \left( \hat{I}_{\pm} \hat{X}_z + \hat{I}_z \hat{X}_{\pm} \right) & \hat{B}_{\pm 1} &= \frac{1}{2} \left( \hat{S}_{\pm} \hat{Y}_z + \hat{S}_z \hat{Y}_{\pm} \right) & F_{\pm 1} &= \pm 3 \sin \theta \cos \theta e^{\mp i\phi} \\ \hat{A}_{\pm 2} &= \frac{1}{2} \hat{I}_{\pm} \hat{X}_{\pm} & \hat{B}_{\pm 2} &= \frac{1}{2} \hat{S}_{\pm} \hat{Y}_{\pm} & F_{\pm 2} &= \pm \frac{3}{2} \sin^2 \theta e^{\mp 2i\phi} \end{aligned}$$

then  $\hat{\Gamma} = \sum_q J_q(\omega_q) \left( \hat{A}_{-q} + \hat{B}_{-q} \right) \left( \hat{A}_q + \hat{B}_q \right)$ , but  $[\hat{A}_q, \hat{B}_q] = 0 \quad \forall q$  hence

$$\hat{\Gamma} = \hat{\Gamma}_A + \hat{\Gamma}_B = \sum_q J_q(\omega_q) \hat{A}_{-q} \hat{A}_q + \sum_q J_q(\omega_q) \hat{B}_{-q} \hat{B}_q$$

and we can just independently sum the resulting relaxation rates.

# Interference Effects

- But what about CSA and C<sub>1</sub>-C<sub>2</sub> dipolar coupling?

$$\hat{H}_{D_{IS}} = -\frac{\gamma_C^2 \hbar}{r_{IS}^3} \sum_{q=2}^2 F_q \hat{A}_q \quad \hat{H}_{CSA1} = \gamma_C B_0 \Delta \sigma \sum_{q=-1}^1 F_q \hat{B}_q \quad \hat{H}_{CSA2} = \gamma_C B_0 \Delta \sigma \sum_{q=-1}^1 F_q \hat{C}_q$$

$$\hat{A}_0 = \sqrt{\frac{1}{6}} \left( 2\hat{I}_z \hat{S}_z - \frac{1}{2}\hat{I}_+ \hat{S}_- - \frac{1}{2}\hat{I}_- \hat{S}_+ \right)$$

$$\hat{B}_0 = \frac{1}{3} \sqrt{\frac{3}{2}} \hat{I}_z$$

$$\hat{C}_0 = \frac{1}{3} \sqrt{\frac{3}{2}} \hat{S}_z$$

$$\hat{A}_{\pm 1} = \frac{1}{2} \left( \hat{I}_{\pm} \hat{S}_z + \hat{I}_z \hat{S}_{\pm} \right)$$

$$\hat{B}_{\pm 1} = \frac{1}{6} \hat{I}_{\pm}$$

$$\hat{C}_{\pm 1} = \frac{1}{6} \hat{S}_{\pm}$$

$$\hat{A}_{\pm 2} = \frac{1}{2} \hat{I}_{\pm} \hat{S}_{\pm}$$

$$F_0 = \sqrt{\frac{3}{2}} (3 \cos^2 \theta - 1)$$

$$F_{\pm 1} = \pm 3 \sin \theta \cos \theta e^{\mp i \phi}$$

$$F_{\pm 2} = \pm \frac{3}{2} \sin^2 \theta e^{\mp 2i \phi}$$

In this case  $\hat{A}_q, \hat{B}_q,$  and  $\hat{C}_q$  don't all commute.  $\hat{\Gamma} = \sum_q J(\omega_q) \left( \hat{A}_{-q} + \hat{B}_{-q} + \hat{C}_{-q} \right) \left( \hat{A}_q + \hat{B}_q + \hat{C}_q \right)$

and we get new cross relaxation terms such as:

$$R_{13} = R_{31} = \langle \hat{I}_z | \hat{\Gamma} | 2\hat{I}_z \hat{S}_z \rangle = \frac{2}{5} q_{CC,CSA1} J(\omega_C) \equiv \frac{1}{T_{Iz-IzS_z-cross}}$$

$$q_{CC,CSA1} = \left( \frac{\mu_0}{4\pi} \right) \frac{\gamma_C^4 \hbar}{r_{CC}^3} B_0 \Delta \sigma_1.$$

$$R_{23} = R_{32} = \langle \hat{S}_z | \hat{\Gamma} | 2\hat{I}_z \hat{S}_z \rangle = \frac{2}{5} q_{CC,CSA2} J(\omega_C) \equiv \frac{1}{T_{S_z-IzS_z-cross}}$$

$$q_{CC,CSA2} = \left( \frac{\mu_0}{4\pi} \right) \frac{\gamma_C^4 \hbar}{r_{CC}^3} B_0 \Delta \sigma_2.$$

# The complete relaxation matrix

$$\frac{d}{dt} \begin{bmatrix} \overline{\langle \hat{I}_z \rangle}(t) \\ \overline{\langle \hat{S}_z \rangle}(t) \\ \overline{\langle 2\hat{I}_z \hat{S}_z \rangle}(t) \end{bmatrix} = - \underbrace{\begin{bmatrix} R_{11} & R_{12} & R_{13} \\ R_{21} & R_{22} & R_{23} \\ R_{31} & R_{32} & R_{33} \end{bmatrix}}_{\text{RELAXATION MATRIX}} \begin{bmatrix} \overline{\langle \hat{I}_z \rangle}(t) - I_z^{eq} \\ \overline{\langle \hat{S}_z \rangle}(t) - S_z^{eq} \\ \overline{\langle 2\hat{I}_z \hat{S}_z \rangle}(t) \end{bmatrix}$$

- Assuming extreme narrowing...

$$R_{11} = \frac{1}{T_{1,CSA_1}} + \frac{1}{T_{1,HC_1}} + \frac{1}{T_{1,CC}}$$

$$R_{22} = \frac{1}{T_{1,CSA_2}} + \frac{1}{T_{1,HC_2}} + \frac{1}{T_{1,CC}}$$

$$R_{33} \approx \frac{1}{T_{1,CSA_1}} + \frac{1}{T_{1,CSA_2}} + \frac{1}{T_{1,HC_1}} + \frac{1}{T_{1,HC_2}} + \frac{2}{5T_{1,CC}}$$

Direct relaxation

$$R_{12} = R_{21} \approx \frac{1}{2T_{1,CC}}$$

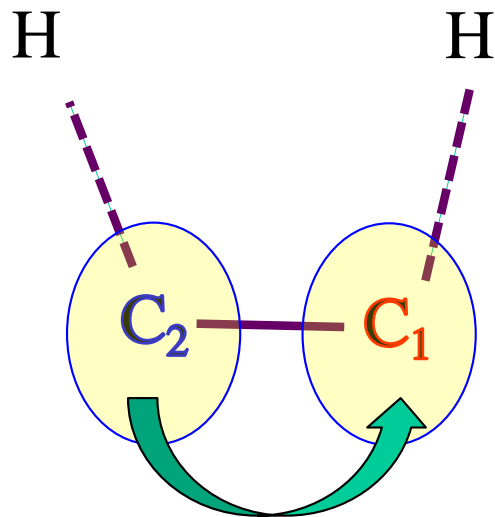
$$R_{13} = R_{31} \approx \sqrt{\frac{6}{5}} \sqrt{\frac{1}{T_{1,CC}T_{1,CSA_1}}}$$

$$R_{23} = R_{32} \approx \sqrt{\frac{6}{5}} \sqrt{\frac{1}{T_{1,CC}T_{1,CSA_2}}}$$

Cross relaxation



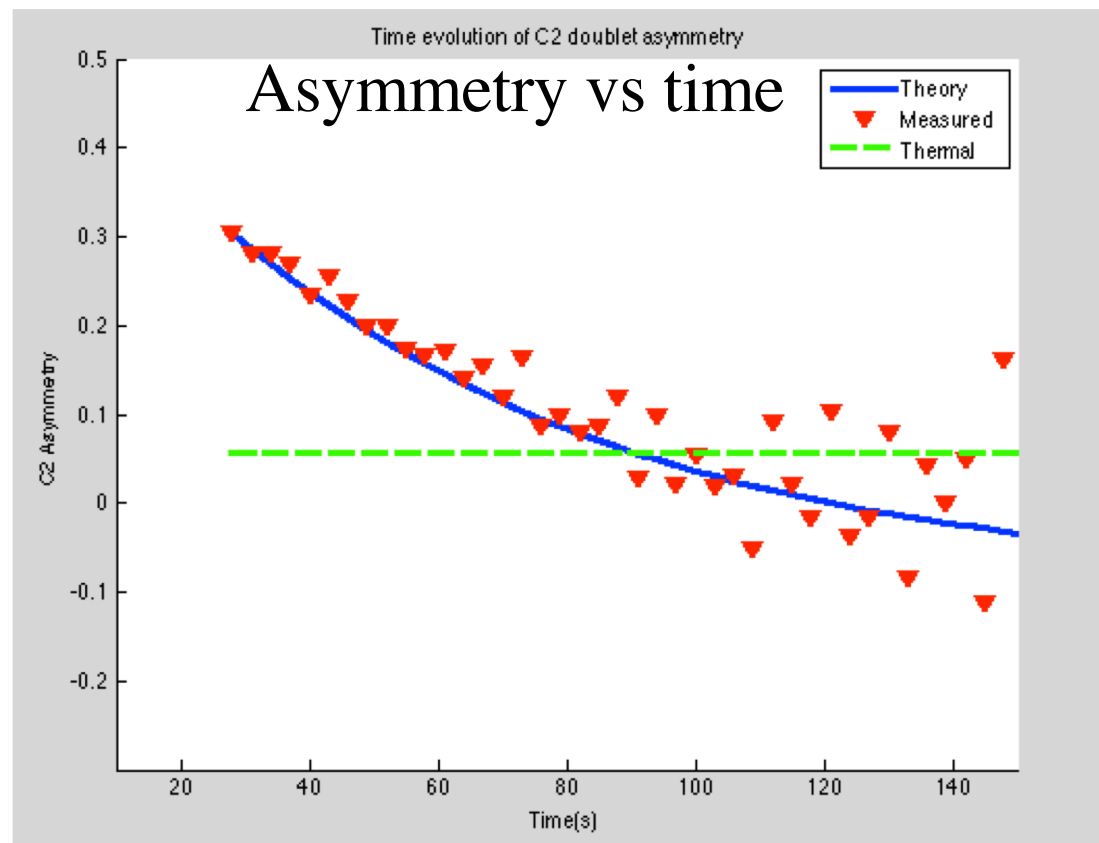
# add Dipolar/CSA Interference



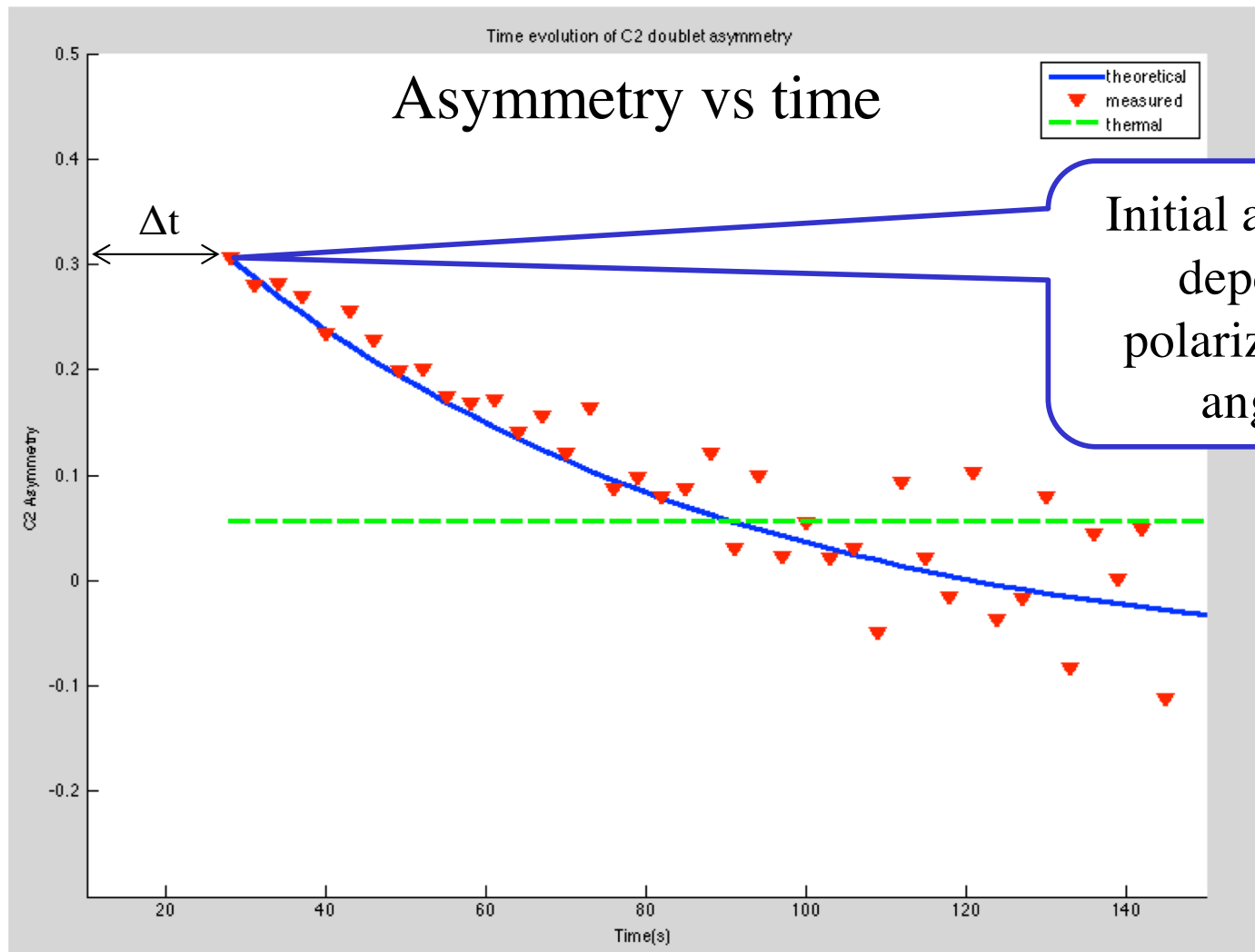
- Despite C-C coupling having a negligible direct effect, cross relaxation (via CSA interference effect) is very important!
- Explains asymmetry beyond equilibrium
- Like CSA, this effect is field dependent.

$$\begin{bmatrix} R_{11} & R_{12} & R_{13} \\ R_{21} & R_{22} & R_{23} \\ R_{31} & R_{32} & R_{33} \end{bmatrix}$$

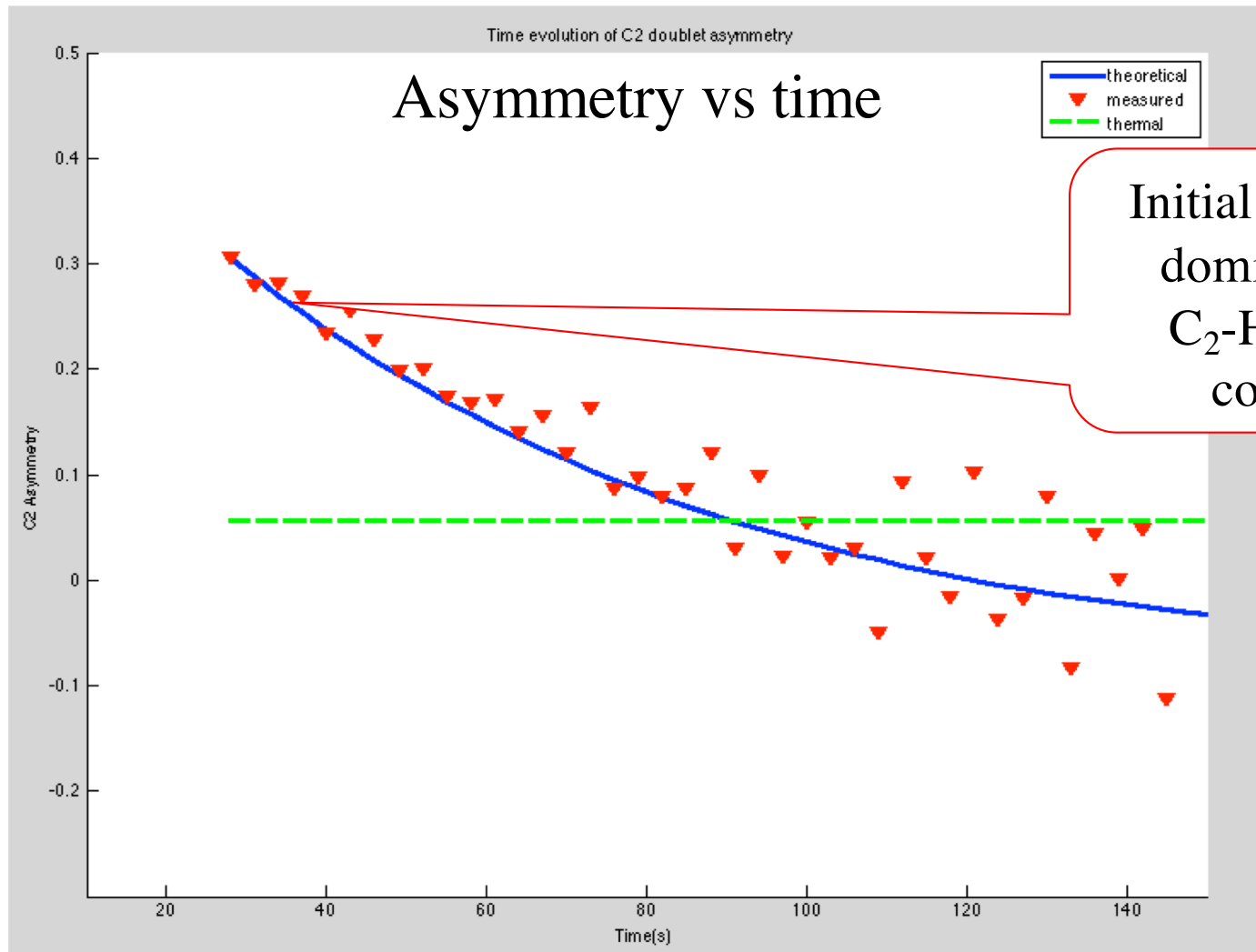
RELAXATION MATRIX



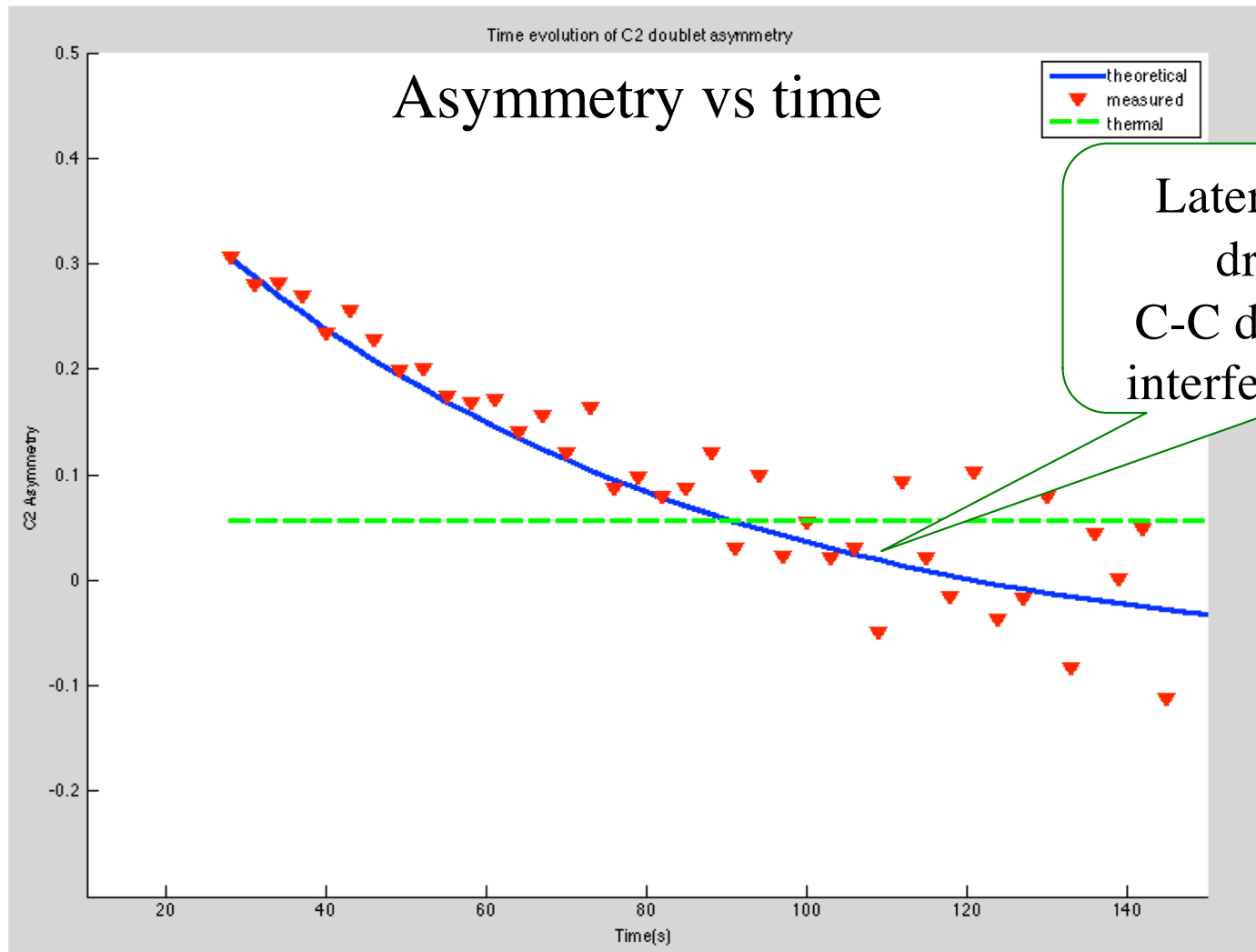
# Doublet asymmetry verses time



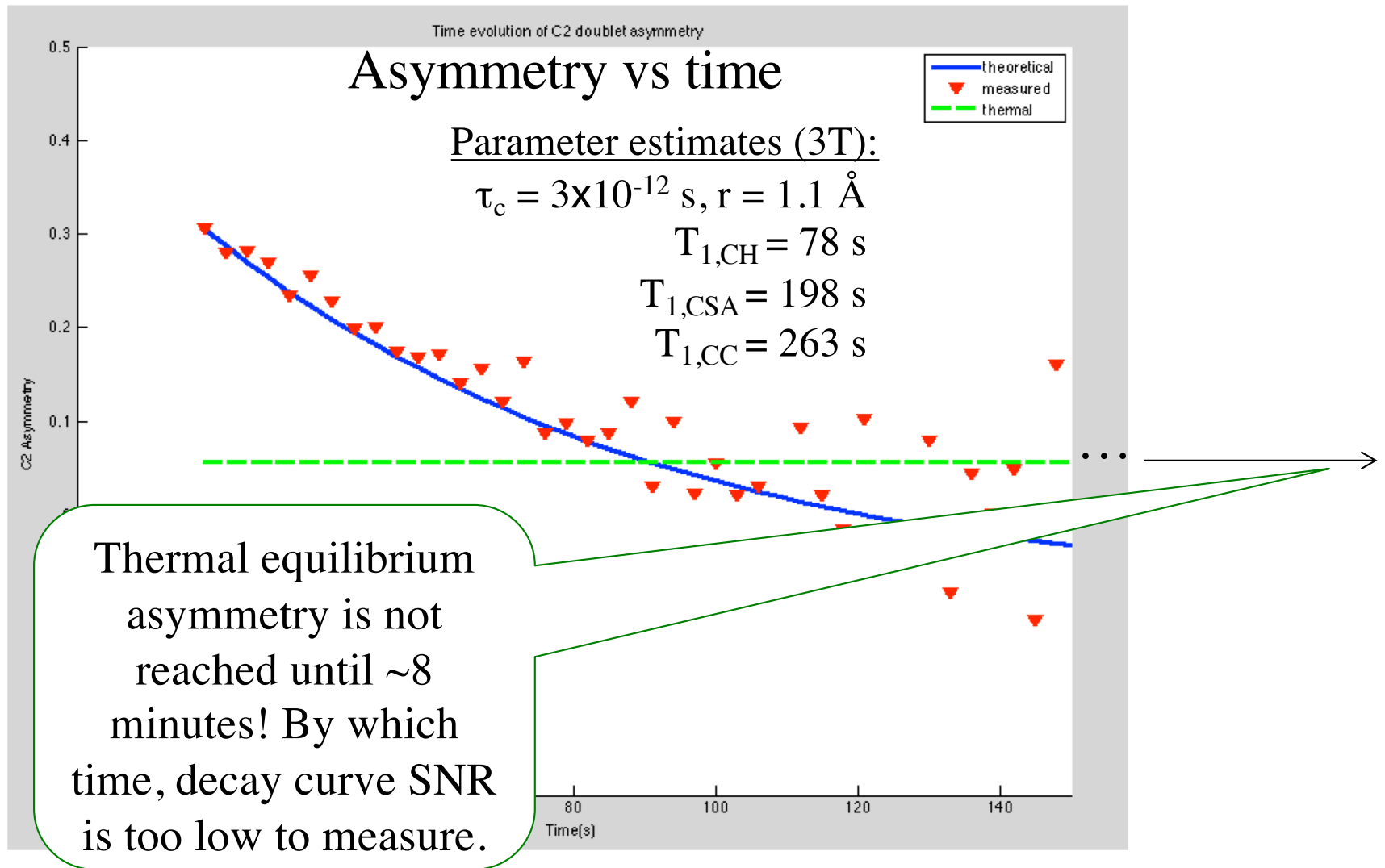
# Doublet asymmetry verses time



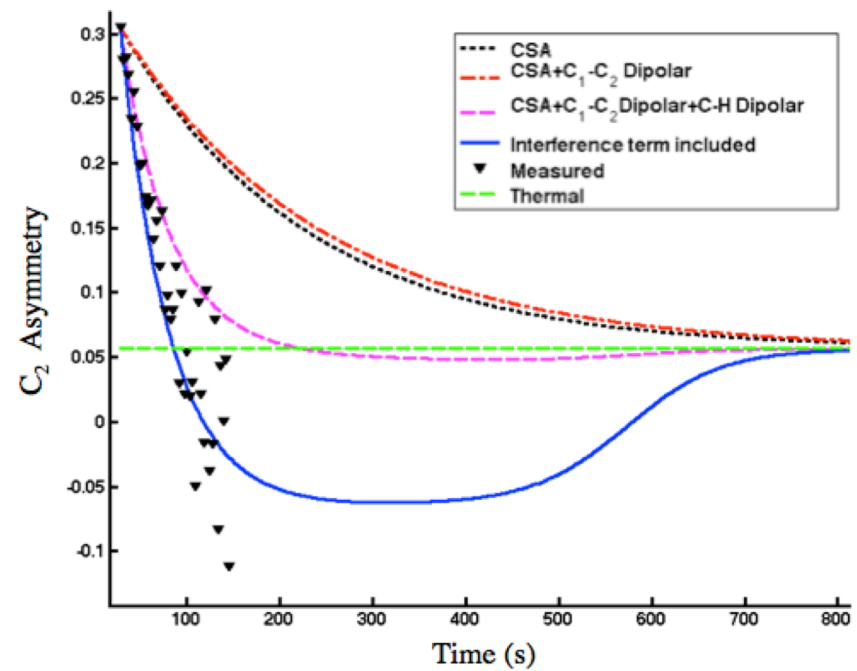
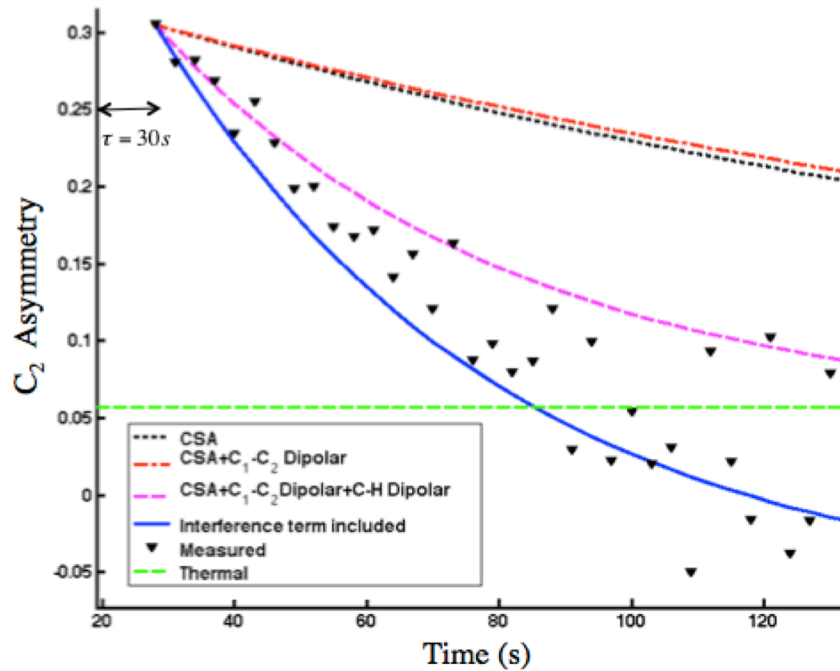
# Doublet asymmetry verses time



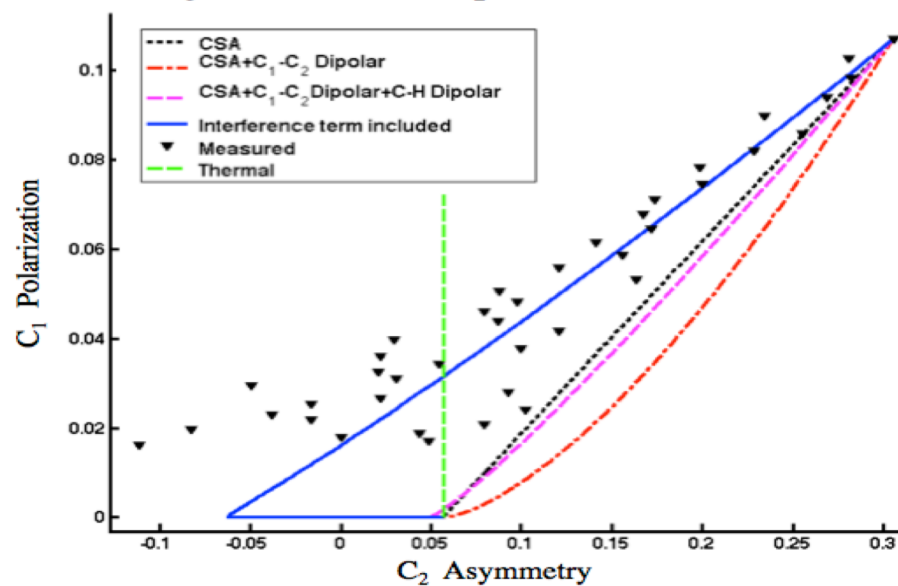
# Doublet asymmetry verses time



# Doublet asymmetry verses time



$C_1$  polarization vs.  $C_2$  doublet asymmetry



# Example 2: Conclusions

- Simple experiment, confusing results.
- Explanation required use of three NMR relaxation mechanisms.
- Asymmetry driven by **three** primary sources.
  - $2I_zS_z$  direct relaxation
  - C-C dipolar coupling/CSA interference effect
  - Residual strong coupling effects
- Use of asymmetry metric to estimate instantaneous polarization requires:
  - Knowledge of initial polarization
  - Time history of the sample in low and high fields

Next Lecture: Spin lattice relaxation  
in the rotating frame ( $T_{1\rho}$ )